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⑦ Applicant: **N.V. INNOGENETICS S.A.**  
**Industriepark Zwijnaarde 7, Box 4**  
**B-9710 Gent(BE)**

⑦ Inventor: **Content, Jean**  
**8 avenue Simone**  
**B-1640 Rhode St-Genèse(BE)**

Inventor: **De Wit, Lucas**  
**Victor Vergauwenstraat 48**  
**B-2870 Puurs(BE)**

Inventor: **De Bruyn, Jacqueline**  
**192 rue de Hongrie**  
**1650 Beersel(BE)**

⑧ Representative: **Grosset-Fournier, Chantal**  
**Catherine et al**  
**ERNEST GUTMANN-YVES PLASSERAUD S.A.,**  
**67 boulevard Haussmann**  
**F-75008 Paris(FR)**

⑨ Polypeptides and peptides, particularly recombinant polypeptides or peptides, nucleic acids coding for the same and use of these polypeptides and peptides in the diagnosis of tuberculosis.

⑩ The invention relates :

- to nucleic acids which contain particularly a nucleotide sequence extending from the extremity constituted by the nucleotide at position (1) to the extremity constituted by the nucleotide at position (149) represented on Figure 1,
- to the polypeptides coded by said nucleic acids.

The polypeptides of the invention can be used for the diagnosis of tuberculosis, and can also be part of the active principle in the preparation of vaccine against tuberculosis.

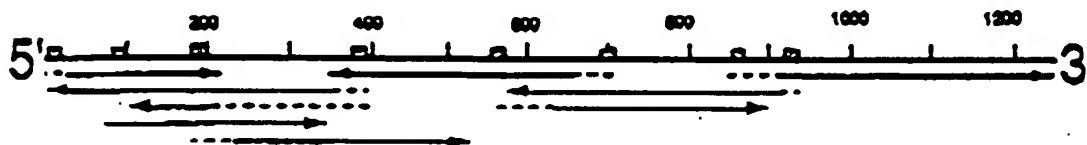


Figure 1

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The invention relates to polypeptides and peptides, particularly recombinant polypeptides and peptides, which can be used for the diagnosis of tuberculosis. The invention also relates to a process for preparing the above-said polypeptides and peptides, which are in a state of biological purity such that they can be used as part of the active principle in the preparation of vaccines against tuberculosis.

It also relates to nucleic acids coding for said polypeptides and peptides.

Furthermore, the invention relates to the *in vitro* diagnostic methods and kits using the above-said polypeptides and peptides and to the vaccines containing the above-said polypeptides and peptides as active principle against tuberculosis.

By "recombinant polypeptides or peptides" it is to be understood that it relates to any molecule having a polypeptidic chain liable to be produced by genetic engineering, through transcription and translation, of a corresponding DNA sequence under the control of appropriate regulation elements within an efficient cellular host. Consequently, the expression "recombinant polypeptides" such as is used herein does not exclude the possibility for the polypeptides to comprise other groups, such as glycosylated groups.

The term "recombinant" indeed involves the fact that the polypeptide has been produced by genetic engineering, particularly because it results from the expression in a cellular host of the corresponding nucleic acid sequences which have previously been introduced into the expression vector used in said host.

Nevertheless, it must be understood that this expression does not exclude the possibility for the polypeptide to be produced by a different process, for instance by classical chemical synthesis according to methods used in the protein synthesis or by proteolytic cleavage of larger molecules.

The expression "biologically pure" or "biological purity" means on the one hand a grade of purity such that the recombinant polypeptide can be used for the production of vaccinating compositions and on the other hand the absence of contaminants, more particularly of natural contaminants.

Tuberculosis remains a major disease in developing countries. The situation is dramatic in some countries, particularly where high incidence of tuberculosis among AIDS patients represents a new source of dissemination of the disease.

Tuberculosis is a chronic infectious disease in which cell-mediated immune mechanisms play an essential role both for protection against and control of the disease.

Despite BCG vaccination, and some effective drugs, tuberculosis remains a major global problem. Skin testing with tuberculin PPD (protein-purified derivative) largely used for screening of the disease is poorly specific, due to cross reactivity with other pathogenic or environmental saprophytic mycobacteria.

Moreover, tuberculin PPD when used in serological tests (ELISA) does not allow to discriminate between patients who have been vaccinated by BCG, or those who have been primo-infected, from those who are developing evolutive tuberculosis and for whom an early and rapid diagnosis would be necessary.

A protein with a molecular weight of 32-kDa has already been purified from zinc deficient *M. bovis* BCG culture filtrate. This protein was identified as antigen 85A (De Bruyn J. et al., 1987, "Purification, partial characterization and identification of a 32-kDa protein antigen of *Mycobacterium bovis* BCG" *Microb. Pathogen.* 2:351). Its NH<sub>2</sub>-terminal amino acid sequence (Phe-Ser-Arg-Pro-Gly-Leu) is identical to that reported for the  $\alpha$ -antigen (antigen 85B) protein purified from *M. bovis* BCG substrain Tokyo (Wiker, H.G. et al., 1988, "MPB59, a widely cross-reacting protein of *Mycobacterium bovis* BCG" *Int. Arch. Allergy Appl. Immunol.* 81:307). The antigen 85-complex is present among different strains of mycobacteria (De Bruyn J. et al., 1989, "Effect of zinc deficiency on the appearance of two immunodominant protein antigens (32-kDa and 65-kDa) in culture filtrates of *Mycobacteria*" *J. Gen. Microbiol.* 135:79). It is secreted by living bacilli as a predominant protein in normal Sauton culture filtrate and could be useful in the serodiagnosis of tuberculosis (Turner M. et al., 1988, "Humoral immune response in human tuberculosis: Immunoglobulins G, A and M directed against the purified P32 protein antigen of *Mycobacterium bovis* bacillus Calmette-Guérin" *J. Clin. Microbiol.* 26:1714) and leprosy (Rumschlag H.B. et al., 1988, "Serological response of patients with lepromatous and tuberculoid leprosy to 30-, 31- and 32-kilodalton antigens of *Mycobacterium tuberculosis*" *J. Clin. Microbiol.* 26:2200). Furthermore, the 32-kDa protein induces specific lymphoproliferation and interferon- $\gamma$  (IFN- $\gamma$ ) production in peripheral blood leucocytes from tuberculosis (Huygen K. et al., 1988, "Specific lymphoproliferation,  $\gamma$ -interferon production and serum immunoglobulin G directed against a purified 32-kDa mycobacterial antigen (P32) in patients with active tuberculosis" *Scand. J. Immunol.* 27:187), and leprosy patients and from PPD- and lepromin-positive healthy subjects. Recent findings indicate that the amount of 32 kDa protein induced IFN- $\gamma$  in BCG-sensitized mouse spleen cells is under probable H-2 control (Huygen K. et al., 1988, "H-2-linked control of *in vitro*  $\gamma$  interferon production in response to a 32-kilodalton antigen (P32) of *Mycobacterium bovis* bacillus Calmette-Guérin" *Infect. Imm.* 56:3188). Finally, the high affinity of mycobacteria for fibronectin is related to proteins of the antigen 85-complex (Abou-Zeid C. et al., 1988, "Characterization of fibronectin-binding antigens released by *Mycobacterium tuberculosis* and *Mycobacterium bovis* BCG" *Infect. Imm.* 56:3046).

Wiker et al. (Wiker H.G. et al., 1990, "Evidence for three separate genes encoding the proteins of the mycobacterial antigen 85 complex" *Infect. Immun.* 58:272) showed recently that the antigens 85A, B and C isolated from *M. bovis* BCG culture filtrate present a few amino acid replacements in their NH<sub>2</sub> terminal region strongly suggesting the existence of multiple genes coding for these proteins. But, the data given for the antigen 85C of *M. bovis* BCG are insufficient to enable its unambiguous identifiability as well as the characterization of its structural and functional elements.

The gene encoding the  $\alpha$ -antigen from *Mycobacterium bovis* BCG has been described (Borremans L. et al., 1988, "Cloning, sequence determination and expression of a 32-Kilodalton protein gene of *Mycobacterium tuberculosis*" *Infect. Immun.* 57:3123) which presented 77.5% homology at the DNA level within the coding region with the  $\alpha$ -antigen gene (Matsuo K. et al., 1988, "Cloning and expression of the *Mycobacterium bovis* BCG gene for extracellular  $\alpha$ -antigen" *J. Bacteriol.* 170:3847). Moreover, we have recently isolated and sequenced a corresponding 32-kDa protein genomic clone from our  $\lambda$ gt11 BCG library (prepared from strain *M. bovis* BCG 1173P2). The complete sequence of this gene is identical with that from *Mycobacterium tuberculosis* except for a single silent nucleotide change (De Wit L. et al., 1990, "Nucleotide sequence of the 32 kDa-protein gene (antigen 85A) of *Mycobacterium bovis* BCG" *Nucl. Ac. Res.* 18:3995). This confirmed the previous findings that partial protein sequence of several tryptic peptides derived from highly purified 32-kDa protein from *M. bovis* BCG present the characteristic 85A sequence (Borremans L. et al., 1988, "Cloning, sequence determination and expression of a 32-Kilodalton protein gene of *Mycobacterium tuberculosis*" *Infect. Immun.* 57:3123 ; De Wit L. et al., 1990, "Nucleotide sequence of the 32 kDa-protein gene (antigen 85A) of *Mycobacterium bovis* BCG" *Nucl. Ac. Res.* 18:3995) and not the 85B sequence. Thus so far it was likely, but not demonstrated, that the genome of *M. bovis* BCG contained at least two genes coding for antigen 85A and 85B respectively. As to the genome of the *Mycobacterium tuberculosis*, nothing was proved as to the existence of new genes, besides the genes coding respectively for 85A and 85B.

An aspect of the invention is to provide with a new family of nucleic acids coding for new proteins and polypeptides which can be used for the detection and control of tuberculosis.

Another aspect of the invention is to provide nucleic acids coding for the peptidic chains of biologically pure recombinant polypeptides which enable their preparation on a large scale.

Another aspect of the invention is to provide antigens which can be used in serological tests as an *in vitro* rapid diagnostic test for tuberculosis.

Another aspect of the invention is to provide a rapid *in vitro* diagnostic means for tuberculosis, enabling it to discriminate between patients suffering from an evolutive tuberculosis from those who have been vaccinated against BCG or who have been primo-infected.

Another aspect of the invention is to provide nucleic probes which can be used as *in vitro* diagnostic reagent for tuberculosis as well as *in vitro* diagnostic reagent for identifying *M. tuberculosis* from other strains of mycobacteria.

The nucleic acids of the invention

- \* contain a nucleotide sequence extending from the extremity constituted by the nucleotide at position (1) to the extremity constituted by the nucleotide at position (148) represented on Figure 1.
- \* or contain one at least of the nucleotide sequences coding for the following peptides or polypeptides :
  - the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (-1) represented on Figure 1, or
  - the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (-1) represented on Figure 1, or
- \* QSSNGQNY, or
- \* PMVQIPRLVA, or
- \* GSTLRNTQTPRDYAADGGRNG or
- \* PPAAPAAPAA.
- \* or contain nucleotidic sequences :
  - hybridizing with the above-mentioned nucleotide sequences, or their complements,
  - complementary to the above-mentioned nucleotide sequences, or
  - which are the above-mentioned nucleotide sequences wherein T can be replaced by U,
- \* or are constituted by the above-mentioned nucleotide sequences.

The hybridization takes place under the following conditions :

- \* hybridization and wash medium :
  - \* a preferred hybridization medium contains about 3 x SSC [SSC = 0.15 M sodium chloride, 0.015 M sodium citrate, pH 7] about 25 mM of phosphate buffer pH 7.1, and 20% deionized formamide, 0.02% Ficoll, 0.02% BSA, 0.02% polyvinylpyrrolidone and about 0.1 mg/ml sheared denatured

salmon sperm DNA.

- a preferred wash medium contains about 3 x SSC, about 25 mM phosphate buffer, pH 7.1 and 20% deionized formamide;

- hybridization temperature (HT) and wash temperature (WT) are between 45°C and 65°C;
- for the nucleotide sequence extending from the extremity constituted by the nucleotide at position (1) to the extremity constituted by the nucleotide at position (149) represented on Figure 1 :

HT = WT = 65°C

for the nucleic acids of the invention defined by coded polypeptides X - Y : i.e.

- the sequence extending from the extremity constituted by the amino acid at position (X) to the extremity constituted by the amino acid at position (Y) represented on Figure 1,

- the sequence extending from the extremity constituted by the amino acid at position (-48) to the extremity constituted by the amino acid at position (-1) represented on Figure 1,

HT = WT = 65°C

- the sequence extending from the extremity constituted by the amino acid at position (-21) to the extremity constituted by the amino acid at position (-1) represented on Figure 1,

HT = WT = 60°C

for the nucleic acids defined by coded polypeptides represented by their sequence :

- SQSNGQNY HT = WT = 45°C

- PMVQIPRLVA HT = WT = 55°C

- GLTLRTNQTPRDTYAAGGRNG HT = WT = 65°C

- PPAAPAAPAA HT = WT = 65°C.

The above-mentioned temperatures are to be expressed as approximately  $\pm 5^\circ\text{C}$ .

Advantageous nucleic acids of the invention contain one at least of the following nucleotide sequences :

- the one extending from the extremity constituted by the nucleotide at position (160) to the extremity constituted by the nucleotide at position (287) on Figure 1,
- the one extending from the extremity constituted by the nucleotide at position (224) to the extremity constituted by the nucleotide at position (287) on Figure 1,
- the one extending from the extremity constituted by the nucleotide at position (537) to the extremity constituted by the nucleotide at position (590) on Figure 1,
- the one extending from the extremity constituted by the nucleotide at position (858) to the extremity constituted by the nucleotide at position (887) on Figure 1,
- the one extending from the extremity constituted by the nucleotide at position (872) to the extremity constituted by the nucleotide at position (1037) on Figure 1,
- the one extending from the extremity constituted by the nucleotide at position (1140) to the extremity constituted by the nucleotide at position (1188) on Figure 1, or contain nucleotidic sequences :
- hybridizing with the above-mentioned nucleotide sequences, or
- complementary to the above-mentioned nucleotide sequences, or
- which are the above-mentioned nucleotide sequences wherein T can be replaced by U, or are constituted by the above-mentioned nucleotide sequences.

The hybridization takes place under the following conditions :

- hybridization and wash medium are as above defined;
  - hybridization temperature (HT) and wash temperature (WT) for the nucleic acids of the invention defined by X - Y : i.e. by the sequence extending from the extremity constituted by the nucleotide at position (X) to the extremity constituted by the nucleotide at position (Y) represented on Figure 1 :
- (160) - (287) HT = WT = 65°C  
 (224) - (287) HT = WT = 60°C  
 (537) - (590) HT = WT = 45°C  
 (858) - (887) HT = WT = 55°C  
 (872) - (1037) HT = WT = 55°C  
 (1140) - (1188) HT = WT = 65°C.

An advantageous group of nucleic acids of the invention contains the nucleotide sequence coding for the following peptide :

SQSNGQNY

and liable to hybridize with the following nucleotide sequence :

CGGCTGGGAC(or T)ATCAACACCCCGGC

and liable to hybridize neither with

GCCTGCGGCAAGGCCGTTGCCAG

nor with

GCCTGCGGTAAGGCTGGCTGCCAG

nor with

GCCTGCGGCAAGGCCGGCTGCACG

or are constituted by the above-mentioned nucleotide sequences.

The above-mentioned hybridization can take place when the hybridization and wash medium is as indicated above; and the hybridization and wash temperature is 52°C.

The expression "not liable to hybridize with" means that the nucleic acid molecule of the invention does not contain a stretch of nucleotide hybridizing at 52°C in the above defined medium with the three probes defined above.

Advantageous nucleic acids of the invention contain one at least of the above-mentioned nucleotide sequences or are constituted by the above-mentioned nucleotide sequences and besides contain an open reading frame coding for a polypeptide

- liable to react selectively with human sera from tuberculosis patients and particularly patients developing an evolutive tuberculosis,
- or liable to be recognized by antibodies also recognizing the amino acid sequence extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (294) represented on Figure 1,
- or liable to generate antibodies recognizing the amino acid sequence extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (294) represented on Figure 1.

The recognition of the above-mentioned sequence of the 294 amino acids (or of the polypeptides of the invention) by the abovesaid antibodies means that the abovesaid sequence forms a complex with one of the above-mentioned antibodies.

Forming a complex between the antigen (i.e. the sequence of 294 amino acids or any polypeptide of the invention) and the antibodies and detecting the existence of a formed complex can be done according to classical techniques (such as the one using a tracer labeled with radioactive isotopes or an enzyme).

Hereafter is given, in a non limitative way, a process for testing the selective reaction between the antigen and human sera from tuberculosis patients and particularly patients developing an evolutive tuberculosis.

This test is an immunoblotting (Western blotting) analysis, in the case where the polypeptides of the invention are obtained by recombinant techniques. This test can also be used for polypeptides of the invention obtained by a different preparation process. After sodium dodecyl sulfate-polyacrylamide gel electrophoresis, polypeptides of the invention are blotted onto nitrocellulose membranes (Hybond c. (Amersham)) as described by Towbin H. et al., 1979, "Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: procedure and some applications" Proc. Natl. Acad. Sci. USA 76:4350-4354. The expression of polypeptides of the invention fused to  $\beta$ -galactosidase in E. coli Y1089, is visualized by the binding of a polyclonal rabbit anti-32-kDa BCG protein serum (1:1,000) or by using a monoclonal anti- $\beta$ -galactosidase antibody (Promega). The secondary antibody (alkaline phosphatase anti-rabbit immunoglobulin G and anti-mouse alkaline phosphatase immunoglobulin G conjugates, respectively) is diluted as recommended by the supplier (Promega).

In order to identify selective recognition of polypeptides of the invention and of fusion proteins of the invention by human tuberculous sera, nitrocellulose sheets are incubated overnight with these sera (1:50) (after blocking a specific protein-binding sites). The human tuberculous sera are selected for their reactivity (high or low) against the purified 32-kDa antigen of BCG tested in a dot blot assay as described in Van Vooren J.P. et al., 1989, "Local anti-P32 humoral response in tuberculous meningitis". Tubercle. 70:123-128. Reactive areas on the nitrocellulose sheets are revealed by incubation with peroxidase conjugated goat anti-human immunoglobulin G antibody (Dakopatts, Copenhagen, Denmark) (1:200) for 4h, and after repeated washings, color reaction is developed by adding peroxidase substrate (o-chloronaphtol) (Bio-Rad Laboratories, Richmond, Calif.) in the presence of peroxidase and hydrogen peroxide.

Advantageous nucleic acids of the invention contain or are constituted by one of the above-mentioned nucleotide sequences, contain an open reading frame and code for a mature polypeptide of about 30 to about 35 kD, and contain a sequence coding for a signal sequence.

Advantageous nucleic acids of the invention contain one at least of the nucleotide sequences coding for the following polypeptides :

- the one extending from the extremity constituted by amino acid at position (-46) to the extremity constituted by amino acid at position (-1) represented on Figure 1, or
- the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (-1) represented on Figure 1, or

- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (284) represented on Figure 1, or
  - the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (284) represented on Figure 1, or
  - the one extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (284) represented on Figure 1,
- or contain nucleotidic sequences :
- hybridizing with the above-mentioned nucleotide sequences, or
  - complementary to the above-mentioned nucleotide sequences, or
  - which are the above-mentioned nucleotide sequences wherein T can be replaced by U, or are constituted by the above-mentioned nucleotide sequences.

The hybridization takes place under the following conditions :

- hybridization and wash medium are as above defined;
  - hybridization temperature (HT) and wash temperature (WT) for the nucleic acids of the invention defined by coded polypeptides X - Y: i.e. by the coded sequence extending from the extremity constituted by the amino acid at position (X) to the extremity constituted by the amino acid at position (Y) represented on Figure 1:
- (-48) - (-1) HT = WT = 85°C  
 (-21) - (-1) HT = WT = 80°C  
 (-48) - (284) HT = WT = 70°C  
 (-21) - (284) HT = WT = 70°C  
 (1) - (284) HT = WT = 70°C.

Advantageous nucleic acids of the invention contain one at least of the following nucleotide sequences :

- the one extending from the extremity constituted by the nucleotide at position (150) to the extremity constituted by the nucleotide at position (287) represented on Figure 1, or
  - the one extending from the extremity constituted by the nucleotide at position (224) to the extremity constituted by the nucleotide at position (287) represented on Figure 1, or
  - the one extending from the extremity constituted by the nucleotide at position (1) to the extremity constituted by the nucleotide at position (1189) represented on Figure 1, or
  - the one extending from the extremity constituted by the nucleotide at position (150) to the extremity constituted by the nucleotide at position (1189) represented on Figure 1, or
  - the one extending from the extremity constituted by the nucleotide at position (224) to the extremity constituted by the nucleotide at position (1189) represented on Figure 1, or
  - the one extending from the extremity constituted by the nucleotide at position (288) to the extremity constituted by the nucleotide at position (1189) represented on Figure 1,
  - the one extending from the extremity constituted by the nucleotide at position (1) to the extremity constituted by the nucleotide at position (1211) represented on Figure 1,
  - the one extending from the extremity constituted by the nucleotide at position (150) to the extremity constituted by the nucleotide at position (1211) represented on Figure 1,
  - the one extending from the extremity constituted by the nucleotide at position (224) to the extremity constituted by the nucleotide at position (1211) represented on Figure 1,
  - the one extending from the extremity constituted by the nucleotide at position (288) to the extremity constituted by the nucleotide at position (1211) represented on Figure 1,
- or contain nucleotidic sequences :
- hybridizing with the above-mentioned nucleotide sequences, or
  - complementary to the above-mentioned nucleotide sequences, or
  - which are the above-mentioned nucleotide sequences wherein T can be replaced by U, or are constituted by one at least of the following nucleotide sequences.

The hybridization takes place under the following conditions :

- hybridization and wash medium are as above defined;
  - hybridization temperature (HT) and wash temperature (WT) for the nucleic acids of the invention defined for the nucleic acids of the invention defined by X - Y: i.e. by the sequence extending from the extremity constituted by the nucleotide at position (X) to the extremity constituted by the nucleotide at position (Y) represented on Figure 1 :
- (150) - (287) HT = WT = 85°C  
 (224) - (287) HT = WT = 80°C  
 (150) - (1189) HT = WT = 70°C  
 (1) - (1189) HT = WT = 70°C

(224) - (1189) HT = WT = 70°C

(288) - (1189) HT = WT = 70°C

An advantageous nucleic acid sequence of the invention contains a nucleotide sequence coding for a polypeptide sequence extending from the extremity constituted by the amino acid at position (18) to the extremity constituted by the amino acid at position (89) represented on Figure 2B, on the fifth line, or is constituted by this nucleotide sequence.

An advantageous nucleic acid sequence of the invention contains a nucleotide sequence constituted by the nucleotide at position (584) to the extremity constituted by the nucleotide at position (828) represented on Figure 2A, on the fifth line, or is constituted by this nucleotide sequence.

The hybridization takes place under the following conditions :

- hybridization and wash medium are as above defined,
- HT = WT = 85°C.

The invention relates also to the polypeptides coded by the nucleic acids of the invention above defined.

Advantageous polypeptides of the invention contain in their polypeptide chain, one at least of the following amino acid sequences :

- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (-1) represented on Figure 1,
- or the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (-1) represented on Figure 1.
- SQSNGQNY,
- PMVQIPRLVA,
- GLTLRTNQTPRDTYAADGGRNG,
- PPAAPAAPAA,

or are constituted by the above-mentioned polypeptide sequences.

Advantageous polypeptides of the invention contain in their polypeptide chain, one at least of the following amino acid sequence :

SQSNGQNY

and the amino acid sequence

GWDINTPA

and containing not the amino acid sequence

ACGKAGCQ

and not the amino acid sequence

ACGKAGCT.

Interesting polypeptides are :

**SQSNGQNY**

**GWDINTPA.**

Advantageous polypeptides of the invention are liable to react selectively with human sera from tuberculosis patients and particularly patients developing an evolutive tuberculosis, or liable to be recognized by antibodies also recognizing the polypeptide sequence extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (284) represented on Figure 1, or liable to generate antibodies recognizing the polypeptide sequence extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (284) represented on Figure 1.

The invention also includes the peptidic sequences resulting from the modification by substitution and/or by addition and/or by deletion of one or several amino acids in the above defined polypeptides and peptides in so far as this modification does not alter the following properties :

selective reaction with human sera from tuberculosis patients and particularly patients developing an evolutive tuberculosis,

and/or reaction with antibodies raised against the amino acid sequence extending from the extremity constituted by amino acid at position (1), to the extremity constituted by amino acid at position (284) represented on Fig. 1.

Advantageous polypeptides of the invention contain or are constituted by one of the above-mentioned polypeptide sequences, and are about 30 to about 35 kD and are preceded by a signal peptide.

Advantageous polypeptides of the invention contain in their polypeptide chain, one at least of the following amino acid sequences or are constituted by one of the following amino acid sequences :

- the one extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (294) represented on Figure 1,
- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (294) represented on Figure 1,
- the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (294) represented on Figure 1,
- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (-1) represented on Figure 1,
- the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (-1) represented on Figure 1.

Advantageous polypeptides of the invention contain in their polypeptide chain the amino acid sequence extending from the extremity constituted by amino acid at position (18) to the extremity constituted by amino acid at position (99) represented on Figure 2B, on the fifth line.

It goes without saying that the free reactive functions which are present in some of the amino acids, which are part of the constitution of the polypeptides of the invention, particularly the free carboxyl groups which are carried by the groups Glu or by the C-terminal amino acid on the one hand and/or the free NH<sub>2</sub> groups carried by the N-terminal amino acid or by amino acid inside the peptidic chain, for instance Lys, on the other hand, can be modified in so far as this modification does not alter the above mentioned properties of the polypeptide.

The molecules which are thus modified are naturally part of the invention. The above mentioned carboxyl groups can be acylated or esterified.

Other modifications are also part of the invention. Particularly, the amine or ester functions or both of terminal amino acids can be themselves involved in the bond with other amino acids. For instance, the N-terminal amino acid can be linked to a sequence comprising from 1 to several amino acids corresponding to a part of the C-terminal region of another peptide.

The polypeptides according to the invention can be glycosylated or not, particularly in some of their glycosylation sites of the type Asn-X-Ser or Asn-X-Thr, X representing any amino acid.

Other advantageous polypeptides of the invention consist in one of the following amino acid sequences :

- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (-1) represented on Figure 1,
- or the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (-1) represented on Figure 1.

These polypeptides can be used as signal peptides, the role of which is to initiate the translocation of a protein from its site of synthesis, but which is excised during translocation.

Advantageous polypeptides of the invention are the ones constituted by :

- SQSNGQNY,
- PMVQIPRLVA,
- GLTLRTNQTPRDTYAADGGRRNG,
- PPAAPAAPAA,
- the one extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (294) represented on Figure 1,
- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (294) represented on Figure 1,
- the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (294) represented on Figure 1,
- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (-1) represented on Figure 1,
- the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (-1) represented on Figure 1,
- GWDINTPA,
- the one extending from the extremity constituted by amino acid at position (18) to the extremity constituted by amino acid at position (99) represented on Figure 2B, on the fifth line.

It is to be noted that the above mentioned polypeptides are derived from the expression products of a DNA derived, as explained hereafter in the examples,

- from the nucleotide sequence coding for a protein of 33-kDa secreted by Mycobacterium tuberculosis



or

- from the nucleotide sequence coding for a protein secreted by *M. bovis* BCG, or
- from related nucleotide sequences which will be hereafter designated by 85-C genes.

The invention also relates to the amino acid sequences constituted by the above mentioned polypeptides and a protein or an heterologous sequence with respect to said polypeptide, said protein or heterologous sequence comprising for instance from about 1 to about 1000 amino acids. These amino acid sequences will be called fusion proteins.

In an advantageous fusion protein of the invention, the heterologous protein is  $\beta$ -galactosidase.

The invention also relates to any recombinant nucleic acids containing at least a nucleic acid of the invention inserted in an heterologous nucleic acid.

The invention relates more particularly to recombinant nucleic acid such as defined, in which the nucleotide sequence of the invention is preceded by a promoter (particularly an inducible promoter) under the control of which the transcription of said sequence is liable to be processed and possibly followed by a sequence coding for transcription termination signals.

The invention also relates to the recombinant nucleic acids in which the nucleic acid sequences coding for the polypeptide of the invention and possibly the signal peptide, are recombined with control elements which are heterologous with respect to the ones to which they are normally associated within the bacteria gene and, more particularly, the regulation elements adapted to control their expression in the cellular host which has been chosen for their production.

The invention also relates to recombinant vectors, particularly for cloning and/or expression, comprising a vector sequence, notably of the type plasmid, cosmid or phage, and a recombinant nucleic acid of the invention, in one of the non essential sites for its replication.

According to an advantageous embodiment of the invention, the recombinant vector contains, in one of its non essential sites for its replication, necessary elements to promote the expression of polypeptides according to the invention in a cellular host and possibly a promoter recognized by the polymerase of the cellular host, particularly an inducible promoter and possibly a signal sequence and/or an anchor sequence.

According to another additional embodiment of the invention, the recombinant vector contains the elements enabling the expression by *E. coli* of a nucleic acid according to the invention inserted in the vector, and particularly the elements enabling the expression of the gene or part thereof of  $\beta$ -galactosidase.

The invention also relates to a cellular host which is transformed by a recombinant vector according to the invention, and comprising the regulation elements enabling the expression of the nucleotide sequence coding for the polypeptide according to the invention in this host.

The invention also relates to a cellular host chosen from among bacteria such as *E. coli*, transformed by a vector as above defined, or chosen from among eukaryotic organism, such as CHO cells, insect cells, Sf9 cells (*Spodoptera frugiperda*) infected by the virus Ac NPV (*Autographa californica* nuclear polyhydrosis virus) containing suitable vectors such as pAc 373 pYM1 or pVC3, BmN [*Bombyx mori*] infected by the virus BmNPV containing suitable vectors such as pBE520 or p89B310.

The invention relates to an expression product of a nucleic acid expressed by a transformed cellular host according to the invention.

The invention also relates to the use of any secreted polypeptide of the invention as a carrier antigen for foreign epitopes (epitopes of a polypeptide sequence heterologous with respect to the polypeptides of the invention) in the *Mycobacterium bovis* BCG vaccine strain.

The *Mycobacterium bovis* BCG vaccine strain used can be available from Institut Pasteur (Paris), under 1173P<sub>2</sub>.

The recombinant DNA comprising the nucleic acid coding for anyone of the polypeptides of the invention and the nucleic acid coding for any foreign epitopes as above defined, can contain the promoter sequence of said polypeptide of the invention, the signal sequence of said polypeptide, possibly the coding part of said polypeptide and the coding nucleic acid of the foreign epitope, said nucleic acid of the foreign epitope being for instance

- either directly located after the signal sequence, and if the coding part of the the polypeptide of the invention is present, upstream the coding part of the polypeptide of the invention,
- or located downstream the coding part of the polypeptide of the invention,
- or located within the coding part of the polypeptide of the invention.

The recombinant DNA as above defined can be transformed into the vaccine strain BCG where it leads to the expression and secretion of a recombinant protein antigen.

From the nucleic acids of the invention, probes (i.e. cloned or synthetic oligonucleotides) can be inferred.

These probes can be from 15 to the maximum number of nucleotides of the selected nucleic acids. The

oligonucleotides can also be used either as amplification primers in the PCR technique (PCR Mullis and Faloona methods in *Enzymology*, vol. 155, p. 335, 1987) to generate specific enzymatically amplified fragments and/or as probes to detect fragments amplified between bracketing oligonucleotide primers.

The specificity of a PCR assisted hybridization assay can be controlled at different levels.

5 The amplification process or the detection process or both can be specific. The latter case giving the higher specificity is preferred.

The invention also relates to a process for preparing a polypeptide according to the invention comprising the following steps:

- the culture in an appropriate medium of a cellular host which has previously been transformed by an appropriate vector containing a nucleic acid according to the invention,
- 10 - the recovery of the polypeptide produced by the above said transformed cellular host from the above said culture medium, and
- the purification of the polypeptide produced, eventually by means of immobilized metal ion affinity chromatography (IMAC).

15 The polypeptides of the invention can be prepared according to the classical techniques in the field of peptide synthesis.

The synthesis can be carried out in homogeneous solution or in solid phase.

For instance, the synthesis technique in homogeneous solution which can be used is the one described by Houbenweyl in the book titled "Methoden der organischen Chemie" (Method of organic chemistry) edited by E. Wunsh, vol. 15-I et II, THIEME, Stuttgart 1974.

20 The polypeptides of the invention can also be prepared according to the method described by R.D. Merrifield in the article titled "Solid phase peptide synthesis" (J.P. Ham.Soc., 45, 2149-2154).

The invention also relates to a process for preparing the nucleic acids according to the invention. A suitable method for chemically preparing the single-stranded nucleic acids (containing at most 100 nucleotides of the invention) comprises the following steps:

- DNA synthesis using the automatic  $\beta$ -cyanoethyl phosphoramidite method described in *Bioorganic Chemistry* 4: 274-325, 1988.

In the case of single-stranded DNA, the material which is obtained at the end of the DNA synthesis can be used as such.

30 A suitable method for chemically preparing the double-stranded nucleic acids (containing at most 100 bp of the invention) comprises the following steps:

- DNA synthesis of one sense oligonucleotide using the automatic  $\beta$ -cyanoethyl phosphoramidite method described in *Bioorganic Chemistry* 4: 274-325, 1988, and DNA synthesis of one anti-sense oligonucleotide using said above-mentioned automatic  $\beta$ -cyanoethyl phosphoramidite method,
- 35 - combining the sense and anti-sense oligonucleotides by hybridization in order to form a DNA duplex,
- cloning the DNA duplex obtained into a suitable plasmid vector and recovery of the DNA according to classical methods, such as restriction enzyme digestion and agarose gel electrophoresis.

A method for the chemical preparation of nucleic acids of length greater than 100 nucleotides - or bp, in the case of double-stranded nucleic acids - comprises the following steps:

- 40 - assembling of chemically synthesized oligonucleotides, provided at their ends with different restriction sites, the sequences of which are compatible with the succession of amino acids in the natural peptide, according to the principle described in *Proc. Nat. Acad. Sci. USA* 80: 7481-7485, 1983,
- cloning the DNA thereby obtained into a suitable plasmid vector and recovery of the desired nucleic acid according to classical methods, such as restriction enzyme digestion and agarose gel electrophoresis.

45 The invention also relates to antibodies themselves formed against the polypeptides according to the invention.

It goes without saying that this production is not limited to polyclonal antibodies.

It also relates to any monoclonal antibody produced by any hybridoma liable to be formed according to classical methods from splenic cells of an animal, particularly of a mouse or rat, immunized against the purified polypeptide of the invention on the one hand, and of cells of a myeloma cell line on the other hand, and to be selected by its ability to produce the monoclonal antibodies recognizing the polypeptide which has been initially used for the immunization of the animals.

55 The invention also relates to any antibody of the invention labeled by an appropriate label of the enzymatic, fluorescent or radioactive type.

The peptides which are advantageously used to produce antibodies, particularly monoclonal antibodies, are the following ones gathered in Table 1 (referring to Figure 1):

Table 1

38	H <sub>2</sub> N-DGLRAQDDYNGWDINTPAFE-COOH	57
78	H <sub>2</sub> N-TDWYQPSQSNQGNYYTYKWT-COOH	97
174	H <sub>2</sub> N-ANSMWCPSSDPAWKRNDFMV-COOH	193
204	H <sub>2</sub> N-RIWVYCGNGTSPDLGGDNIP-COOH	223
235	H <sub>2</sub> N-NQTFRDITYAADGGRNGVFHF-COOH	254
250	H <sub>2</sub> N-GVFNFPPNGTHSWPYWNEQL-COOH	269
275	H <sub>2</sub> N-DIQHVLNGATPPAAPAAPAA-COOH	294

The amino acid sequences are given in the 1-letter code.

Variations of the peptides listed in Table 1 are also possible depending on their intended use. For example, if the peptides are to be used to raise antisera, the peptides may be synthesized with an extra cysteine residue added. This extra cysteine residue is preferably added to the amino terminus and facilitates the coupling of the peptide to a carrier protein which is necessary to render the small peptide immunogenic. If the peptide is to be labeled for use in radioimmune assays, it may be advantageous to synthesize the protein with a tyrosine attached to either the amino or carboxyl terminus to facilitate iodination. These peptides possess therefore the primary sequence of the peptides listed in Table 1 but with additional amino acids which do not appear in the primary sequence of the protein and whose sole function is to confer the desired chemical properties to the peptides.

The invention also relates to a process for detecting in vitro antibodies related to tuberculosis in a human biological sample liable to contain them, this process comprising

- contacting the biological sample with a polypeptide or a peptide according to the invention under conditions enabling an in vitro immunological reaction between said polypeptide and the antibodies which are possibly present in the biological sample and
- the in vitro detection of the antigen/antibody complex which may be formed.

Preferably, the biological medium is constituted by a human serum.

The detection can be carried out according to any classical process.

By way of example a preferred method brings into play an immunoenzymatic process according to ELISA technique or immunofluorescent or radioimmunological (RIA) or the equivalent ones.

Thus the invention also relates to any polypeptide according to the invention labeled by an appropriate label of the enzymatic, fluorescent, radioactive... type.

Such a method for detecting in vitro antibodies related to tuberculosis comprises for instance the following steps:

- deposit of determined amounts of a polypeptidic composition according to the invention in the wells of a titration microplate,
- introduction into said wells of increasing dilutions of the serum to be diagnosed,
- incubation of the microplate,
- repeated rinsing of the microplate,
- introduction into the wells of the microplate of labeled antibodies against the blood immunoglobulins,
- the labeling of these antibodies being carried out by means of an enzyme which is selected from among the ones which are able to hydrolyze a substrate by modifying the absorption of the radiation of this latter at least at a given wave length,
- detection by comparing with a control standard of the amount of hydrolyzed substrate.

The invention also relates to a process for detecting and identifying in vitro antigens of M. tuberculosis in a human biological sample liable to contain them, this process comprising: - contacting the biological sample with an appropriate antibody of the invention under conditions enabling an in vitro immunological reaction between said antibody and the antigens of M. tuberculosis which are possibly present in the biological sample and the in vitro detection of the antigen/antibody complex which may be formed.

Preferably, the biological medium is constituted by sputum, pleural effusion liquid, broncho-alveolar washing liquid, urine, biopsy or autopsy material.

Appropriate antibodies are advantageously monoclonal antibodies directed against the peptides which have been mentioned in Table 1.

The invention also relates to an additional method for the in vitro diagnosis of tuberculosis in a patient liable to be infected by Mycobacterium tuberculosis comprising the following steps:

- the possible previous amplification of the amount of the nucleotide sequences according to the invention, liable to be contained in a biological sample taken from said patient by means of a DNA primer set as above defined,
- contacting the above mentioned biological sample with a nucleotide probe of the invention, under conditions enabling the production of an hybridization complex formed between said probe and said nucleotide sequence,
- detecting the above said hybridization complex which has possibly been formed.

To carry out the in vitro diagnostic method for tuberculosis in a patient liable to be infected by Mycobacterium tuberculosis as above defined, the following necessary or kit can be used, said necessary or kit comprising:

- a determined amount of a nucleotide probe of the invention,
- advantageously the appropriate medium for creating an hybridization reaction between the sequence to be detected and the above mentioned probe,
- advantageously, reagents enabling the detection of the hybridization complex which has been formed between the nucleotide sequence and the probe during the hybridization reaction.

The invention also relates to an additional method for the in vitro diagnosis of tuberculosis in a patient liable to be infected by Mycobacterium tuberculosis comprising:

- contacting a biological sample taken from a patient with a polypeptide or a peptide of the invention, under conditions enabling an in vitro immunological reaction between said polypeptide or peptide and the antibodies which are possibly present in the biological sample and
- the in vitro detection of the antigen/antibody complex which has possibly been formed.

To carry out the in vitro diagnostic method for tuberculosis in a patient liable to be infected by Mycobacterium tuberculosis, the following necessary or kit can be used, said necessary or kit comprising:

- a polypeptide or a peptide according to the invention,
- reagents for making a medium appropriate for the immunological reaction to occur,
- reagents enabling to detect the antigen/antibody complex which has been produced by the immunological reaction, said reagents possibly having a label, or being liable to be recognized by a labeled reagent, more particularly in the case where the above mentioned polypeptide or peptide is not labeled.

The invention also relates to an additional method for the in vitro diagnosis of tuberculosis in a patient liable to be infected by M. tuberculosis, comprising the following steps:

- contacting the biological sample with an appropriate antibody of the invention under conditions enabling an in vitro immunological reaction between said antibody and the antigens of M. tuberculosis which are possibly present in the biological sample and - the in vitro detection of the antigen/antibody complex which may be formed.

Appropriate antibodies are advantageously monoclonal antibodies directed against the peptides which have been mentioned in Table 1.

To carry out the in vitro diagnostic method for tuberculosis in a patient liable to be infected by Mycobacterium tuberculosis, the following necessary or kit can be used, said necessary or kit comprising:

- an antibody of the invention,
- reagents for making a medium appropriate for the immunological reaction to occur,
- reagents enabling to detect the antigen/antibody complexes which have been produced by the immunological reaction, said reagent possibly having a label or being liable to be recognized by a label reagent, more particularly in the case where the above mentioned antibody is not labeled.

An advantageous kit for the in vitro diagnosis of tuberculosis comprises:

- at least a suitable solid phase system, e.g. a microtiter-plate for deposition thereon of the biological sample to be diagnosed in vitro,
- a preparation containing one of the monoclonal antibodies of the invention,
- a specific detection system for said monoclonal antibody,
- appropriate buffer solutions for carrying out the immunological reaction between a test sample and said monoclonal antibody on the one hand, and the bonded monoclonal antibodies and the detection system on the other hand.

The invention also relates to a kit, as described above, also containing a preparation of one of the polypeptides or peptides of the invention, said antigen of the invention being either a standard (for quantitative determination of the antigen of M. tuberculosis which is sought) or a competitor, with respect to the antigen which is sought, for the kit to be used in a competition dosage process.

The invention also relates to an immunogenic composition comprising a polypeptide or a peptide according to the invention, in association with a pharmaceutically acceptable vehicle.

The invention also relates to a vaccine composition comprising among other immunogenic principles anyone of the polypeptides or peptides of the invention or the expression product of the invention, possibly coupled to a natural protein or to a synthetic polypeptide having a sufficient molecular weight so that the conjugate is able to induce in vivo the production of antibodies neutralizing Mycobacterium tuberculosis, or induce in vivo a cellular immune response by activating M. tuberculosis antigen-responsive T cells.

The peptides of the invention which are advantageously used as immunogenic principle are the ones mentioned in Table 1.

Other characteristics and advantages of the invention will appear in the following examples and the figures illustrating the invention.

## FIGURE LEGENDS

### Figure 1 :

Figure 1 represents the nucleotide and amino acid sequence of the 85-O antigen containing region of M. tuberculosis :

The previously identified 28 residue NH<sub>2</sub>-terminal amino acid sequence of the mature protein is underlined with a double line. One additional ATG codon, downstream of the ATG at position 150 is underlined. Since the precise length of the signal sequence could not be determined, the option taken here represents the 48 amino acid signal peptide corresponding to ATG<sub>150</sub>. The putative signal peptide sequence is represented in *italic capitals*. The top drawing represents the sequencing strategy. Arrows indicate the direction of dideoxy-sequencing either in DNA subcloned as double stranded DNA in Blue Scribe M13+ or as single stranded DNA in the mp18 M13 vector. The entire sequence was determined using synthetic oligonucleotides represented as gray boxes on the figure.

### Figure 2 :

Figure 2 represents the homology between known nucleotide and amino acid sequence of the antigen 85 and the 85-C antigen of M. tuberculosis :

A- Comparison of the DNA sequences of antigen 85-A, B and C :

DNA sequences have been aligned with the "Align" program which visualizes multiple alignments. In this presentation, sequence differences are outlined :

(•) indicate identical residues ; (-) indicates a gap ; (any letter) indicates a substitution.

All the sequences are compared and aligned to that of the first line (gene 85-A).

85-A : DNA sequence from M. tuberculosis (Borremans L. et al., 1989, "Cloning, sequence determination and expression of a 32-Kilodalton protein gene of Mycobacterium tuberculosis" Infect. Immun. 57:3123).

85-B : DNA sequence from  $\alpha$ -antigen of Mycobacterium bovis (strain Tokyo)(Matsuo K. et al., 1988, "Cloning and expression of the Mycobacterium bovis BCG gene for extracellular  $\alpha$ -antigen" J. Bacteriol. 170:3847).

85-C : DNA sequence from antigen 85-C (the present invention).

85-B-Kan : DNA sequence from antigen 85-B from M. kansasii (Matsuo K. et al., 1990, "Cloning and expression of the gene for cross-reactive  $\alpha$  antigen of Mycobacterium kansasii" Infect. Immun. 58:550).

85-C-BCG : DNA sequence from Mycobacterium bovis BCG strain 1173P2 (the present invention).

Indicates the presumed initiation codon for each gene.

(i) indicates the first phenylalanine residue of the mature protein.



Indicates the termination codon of each gene.

P78 and P79 are sense and antisense primers used for PCR amplification

85-A, -B, -C sequences used for the synthesis of specific synthetic oligonucleotide probes are framed.

The indicated restriction sites have been used to prepare the three type specific probes.

B- Comparison of the Pre-protein sequences of antigen 85-A, B and C :

DNA sequences have been aligned with the "Align" program which permits multiple alignments. In this presentation, sequence differences are outlined :

(•) indicate identical residues ; (-) indicates a gap ; (any letter) indicates a substitution.

All the sequences are compared and aligned to that of the first line (gene 85-A).

85-A : Protein sequence from *M. tuberculosis* (Borremans L. et al., 1989, "Cloning, sequence determination and expression of a 32-Kilodalton protein gene of *Mycobacterium tuberculosis*" Infect. Immun. 57:3123).

85-B : Protein sequence from  $\alpha$ -antigen of *Mycobacterium bovis* (strain Tokyo) (Matsuo K. et al., 1988, "Cloning and expression of the *Mycobacterium bovis* BCG gene for extracellular  $\alpha$ -antigen" J. Bacteriol. 170:3847).

85-C : Protein sequence from antigen 85-C (the present invention).

85-B-Kans : Protein sequence from antigen 85-B from *M. kansasii* (Matsuo K. et al., 1990, "Cloning and expression of the gene for cross-reactive  $\alpha$  antigen of *Mycobacterium kansasii*" Infect. Immun. 58:550).

85-C-BCG : Protein sequence from *Mycobacterium bovis* BCG strain 1173P2 (the present invention).

The "C" characteristic motif is framed.

Figure 3 :

Figure 3 represents the hydropathy pattern of the *M. tuberculosis* 32-kDa (antigen 85-A), the  $\alpha$ -antigen of BCG (antigen 85-B) and antigen 85-C from *M. tuberculosis*, amino acid sequences :

The sequence of the three pre-proteins (including the presumed signal peptide signals) have been analyzed using the Kyte and Doolittle method (Borremans L. et al., 1989, "Cloning, sequence determination and expression of a 32-Kilodalton protein gene of *Mycobacterium tuberculosis*" Infect. Immun. 57:3123) with a window of eight amino acids. Each bar on the axes represents 50 amino acids. Since the length of signal sequences are slightly different (43, 40 and 46 residues for the three proteins 85-A, 85-B, 85-C) the patterns are aligned to the first residue of the three mature proteins. Plain lines are used to align hydrophobic peaks and a dashed line to align hydrophilic peaks.

Figures 4A and 4B :

Figure 4A represents the restriction endonuclease maps of the three genes 85-A, 85-B and 85-C :

The map of gene 85-A is derived from Borr et al. (Borremans L. et al., 1989, "Cloning, sequence determination and expression of a 32-Kilodalton protein gene of *Mycobacterium tuberculosis*" Infect. Immun. 57:3123). The map of 85-B was obtained from clone 5.1 derived from our *Mycobacterium bovis* BCG 1173P2  $\lambda$ gt11 recombinant library (De Wit L. et al., 1990, "Nucleotide sequence of the 32 kDa-protein gene (antigen 85A) of *Mycobacterium bovis* BCG" Nucl. Ac. Res. 18:3995). For the restriction enzymes used, this map is identical to that published for *M. bovis* BCG (strain Tokyo) (Matsuo K. et al., 1988, "Cloning and expression of the *Mycobacterium bovis* BCG gene for extracellular  $\alpha$ -antigen" J. Bacteriol. 170:3847). The coding region of the 85-B antigen is positioned according to Matsuo et al. (Matsuo K. et al., 1988, "Cloning and expression of the *Mycobacterium bovis* BCG gene for extracellular  $\alpha$ -antigen" J. Bacteriol. 170:3847).

The map of 85-C corresponds to the restriction map of clone 11.2 that was obtained from the *M. tuberculosis*  $\lambda$ gt11 library from R. Young (Young R.A. et al., 1985, "Dissection of *Mycobacterium tuberculosis* antigens using recombinant DNA" Proc. Natl. Acad. Sci. USA 82:2683) (Material and Methods). The position of the specific 5' DNA restriction fragment used for Southern analysis is indicated on each map, by a double arrow.

Figure 4B represents the Southern analysis of the total genomic DNA from *Mycobacterium bovis* BCG - (strain 1173P2) :

15  $\mu$ g DNA of digested DNA was applied per lane. Hybridization was either with oligonucleotide probes A, B, C (as described in Fig. 2A) or, after deahybridization, with a larger DNA fragment in conditions described in Material and methods. Part 85-C was obtained on a separate gel. Molecular weights of the hybridizing bands were calculated by comparison with standards.

Figure 5 :

Figure 3 represents the pulse field electrophoresis of *Mycobacterium tuberculosis* DNA :

DNA from three strains of *Mycobacterium tuberculosis* was digested with DraI and separated by Pulse field electrophoresis on an agarose gel together with a bacteriophage  $\lambda$  DNA 'ladder' as described in Material and methods. After transfer to Nylon filters, hybridization with the three probes 85-A, 85-B, 85-C was as described under Fig. 4. Molecular weights of the hybridizing bands were calculated by comparison with those of the  $\lambda$  DNA 'ladder'.

## MATERIAL AND METHODS

1. Preparation of genomic DNA (Thole J. et al., 1986, "Cloning of *Mycobacterium bovis* BCG DNA and expression of antigens in *Escherichia coli*" Infect. Immun. 50:3800) :

*M. bovis* BCG was cultivated at 37°C in Sauton medium and harvested after an additional incubation of 18 hrs in the presence of 1% glycine added at the end of the late exponential growth phase. The bacteria were treated with lysozyme and proteinase K, lysed with sodium dodecyl sulfate, phenol extracted and ethanol precipitated.

### 2. Genomic libraries :

A  $\lambda$ gt11 recombinant library constructed from genomic DNA of *M. tuberculosis* (Erdman strain), was obtained from Young R.A. et al., 1985, "Dissection of *Mycobacterium tuberculosis* antigens using recombinant DNA" Proc. Natl. Acad. Sci. USA 82:2683.

A second  $\lambda$ gt11 recombinant library was prepared with genomic DNA from *M. bovis* BCG (De Wit L. et al., 1990, "Nucleotide sequence of the 32 kDa-protein gene (antigen 85A) of *Mycobacterium bovis* BCG" Nucl. Ac. Res. 18:3995).

### 3. Oligonucleotides :

Oligonucleotides were synthesized on an Applied Biosystems DNA synthesizer model 381A, purified on OPC-cartridges (Applied Biosystems), lyophilized and dissolved in TE buffer (10 mM Tris-HCl, pH 7.4).

<sup>32</sup>P labelling of the oligonucleotides was as described in Sambrook J. et al., 1989, "Molecular cloning: a laboratory manual" Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.

### 4. PCR :

#### 4.1. Amplification and cloning of genomic DNA:

50 ng of *Mycobacterium bovis* BCG DNA was amplified in a 50  $\mu$ l reaction containing 1xPCR-buffer (Amersham), 200  $\mu$ M dNTP, 1  $\mu$ M each of sense P78 (5'-CCGGAATTCATGGGCCGTGACATCAAG) and antisense P79 (5'-CCGGAATTCGGTCTCCCACTTGTAAGT) oligonucleotide primers (the location of these two primers is indicated in Figure 2A. Both oligonucleotides were added an EcoRI sequence preceded by 3 additional nucleotides), and 2 units of Tag DNA polymerase. After denaturation for 90 seconds at 94°C the reaction was submitted to 40 cycles consisting of 1 minute at 93°C (denaturation), 90 seconds at 55°C (annealing), 2 minutes at 72°C (extension), followed by a 6 minutes final extension at 72°C. After extraction with 150  $\mu$ l chloroform, the amplified DNA, was washed three times with 0.75 ml H<sub>2</sub>O in a Centricon-30 for 6 minutes at 8500 rpm in the Sorvall SS 34 rotor. After digestion with EcoRI the DNA was ligated into EcoRI-digested, phosphatase-treated Bluescribe-M13+ vector, DH5 $\alpha$  *E. coli* (Gibco-BRL) were transformed and plated on Hybond-N filters. Colonies were selected by hybridization with <sup>32</sup>P-oligonucleotide probe-A (5'-TOGCCCCGCCCTGTACCTG) and probe-B (5'-TCACCTGCGGTTTATCTG). Hybridization and washing conditions for the oligonucleotides were as described by Jacobs et al. (Jacobs et al., 1988, "The thermal stability of oligonucleotide duplexes is sequence independent in tetraalkylammonium salt solutions: application to identifying recombinant DNA clones" Nucl. Ac. Res. 16:4837).

#### 4.2. Amplification of $\lambda$ gt11 plaques DNA :

10  $\mu$ l of each  $\lambda$ gt11 plaque (1 plaque was resuspended in 1 ml 8M medium containing 5% chloroform) were amplified in a 100  $\mu$ l reaction containing 200  $\mu$ M of each dNTP, 1  $\mu$ M each of sense oligonucleotide B and antisense oligonucleotide P79 primers, and 1x PCR buffer (Amersham). After an initial denaturation for



90 seconds at 94°C, 2 units of Taq DNA polymerase were added. The reaction was submitted to 40 cycles consisting of 1 minute at 94°C (denaturation), 90 seconds at 60°C (annealing), 2 minutes at 72°C (extension) followed by a 5 minutes final extension at 72°C. 10 µl were analyzed on a 2% agarose gel stained with ethidium bromide.

#### 5. Screening of the *λ*gt11 *M. tuberculosis* and *Mycobacterium bovis* BCG recombinant DNA libraries:

The two *λ*gt11 recombinant libraries were screened by colony hybridization (Sambrook J. et al., 1989, "Molecular cloning: a laboratory manual" Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.) with a 800 bp Hind III fragment which does not discriminate gene 85-A from 85-B (see Fig. 2A). 12 positive *M. tuberculosis* and 12 *Mycobacterium bovis* BCG plaques were retained and screened by PCR-amplification with primers B (sense) and P78 (antisense). Non-A recombinant *λ*gt11 recombinants were further screened by hybridization with <sup>32</sup>P-labeled-C oligonucleotide-probe (5'-TCGCAGAGCAACGGGCCAGAACTAC) as described above.

From the *M. tuberculosis* *λ*gt11 library, one selected bacteriophage #11 was partially digested with EcoRI and its 5 kbp insert was subcloned in Bluescribe-M13+. From this recombinant plasmid named 11-2, a 3,500 bp BamHI-EcoRI fragment was subcloned in M13-mp18 and M13-mp19 (Sambrook J. et al., 1989, "Molecular cloning: a laboratory manual" Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.).

#### 6. Recombinant DNA analysis:

It was as described in Borremans L. et al., 1989, "Cloning, sequence determination and expression of a 32-Kilodalton protein gene of *Mycobacterium tuberculosis*" Infect. Immun. 57:3123.

#### 7. Sequencing:

Sequence analysis was done by the primer extension dideoxy termination method of Sanger et al. (Sanger F. et al., 1977, "DNA sequencing with chain termination inhibitors" Proc. Natl. Acad. Sci. USA 74:5463) after subcloning of specific fragments in Bluescribe-M13+ (Chen E.J. et al., 1985, "Supercoll sequencing: a fast simple method for sequencing plasmid DNA" DNA 4:166) or in mp18 and mp19 M13 vectors. Sequence analysis was greatly hampered by the high GC content of the *M. tuberculosis* DNA (85%). Sequencing reactions were therefore performed with several DNA polymerases according to manufacturers protocols: T7 DNA polymerase ("Sequenase" USB), T7 DNA polymerase (Pharmacia), and Taq DNA polymerase (Promega) using 7-deaza-dGTP instead of dGTP. Several oligodeoxynucleotides were synthesized and used to focus on ambiguous regions of the sequence. The sequencing strategy is summarized in Fig. 1.

#### 8. Sequence comparison and analysis:

Routine computer aided analysis of the nucleic acid and deduced amino acid sequences were performed with the LGBC program from Ballon B., 1988, "Apple Macintosh programs for nucleic and protein sequence analysis" Nucleic Acid Res. 16:1837. Homology searches used the FASTA programs from Pearson W.R. et al., 1988, "Improved tools for biological sequence comparison" Proc. Natl. Acad. Sci. USA 85:2444, and the various DNA and protein data bank from the EMBL-server facilities. Multiple alignments were obtained with 'Align 1.01' (Scientific and Educational Software).

#### 9. Southern blot analysis:

Genomic DNA from *Mycobacterium bovis* BCG was completely digested with SphI, EcoRI or KpnI, electrophoresed on a 1% agarose gel, transferred to Hybond-N filter (Amersham) after denaturation and neutralization and either hybridized with <sup>32</sup>P-labeled-oligonucleotide probes (A, B, C) in the conditions described in Jacobs et al., 1988, "The thermal stability of oligonucleotide duplexes is sequence independent in tetraalkylammonium salt solutions: application to identifying recombinant DNA clones" Nucl. Ac. Res. 16:4837, or random-primed <sup>32</sup>P-labeled DNA restriction fragments that were found to discriminate the 3 genes 85-A, 85-B, and 85-C.

Probes 85-A was a 230 bp PstI fragment from plasmid BY-5 (Borremans L. et al., 1989, "Cloning, sequence determination and expression of a 32-Kilodalton protein gene of *Mycobacterium tuberculosis*" Infect. Immun. 57:3123 and Fig. 2A). Probes 85-B was a 400 bp SmaI-EcoRV fragment from a 85-B



recombinant plasmid named 5.1, derived from our *Mycobacterium bovis* BCG  $\lambda$ gt11 library, whose map is presented in Fig. 4A (see also Fig. 2A). Probe 85-C was a 280 bp *Sma*I-*Kpn*I fragment from plasmid 11.2 (see also Fig. 4A and 2A).

- These DNA fragments were prepared by gel electrophoresis on low melting point agarose followed by a rapid purification on QIAGEN (marketed by : Westburg, Netherlands) (Up 5) according to manufacturers protocol and labeled in the presence of  $\alpha$ - $^{32}$ P-dCTP (Feinberg A.P. et al., 1983, "A technique for radiolabelling DNA restriction endonuclease fragments to high specific activity" *Anal. Biochem.* **132**:6).

#### 10. Pulse Field electrophoresis DNA separation :

- DNA preparation, restriction enzyme digestion and pulse-field gel electrophoresis were performed as previously described (Vincent Levy-Frebault V. et al., 1990, "DNA polymorphism in *Mycobacterium paratuberculosis*, "wood pigeon mycobacteria" and related mycobacteria analyzed by field inversion gel electrophoresis", *J. Clin. Microbiol.* **27**:2723). Briefly cells from fresh cultures were mixed with 1% low-melting-point agarose (v/v) and submitted to successive treatments with zymolase (Seikagaki Kogyo, Tokyo, Japan), lysozyme, and sodium dodecyl sulfate in the presence of proteinase K (Boehringer GmbH, Mannheim, Germany). After inactivation of proteinase K with phenylmethylsulfonyl fluoride (Bio-Rad Laboratories), agarose blocks were digested overnight with 50 U of *Dra*I (Bio-Rad Laboratories). Then blocks were loaded into a 1% agarose gel prepared and electrophoresed in 0.68 TBE (Tris-boric acid - EDTA) (Vincent Levy-Frebault V. et al., 1990, "DNA polymorphism in *Mycobacterium paratuberculosis*, "wood pigeon mycobacteria" and related mycobacteria analyzed by field inversion gel electrophoresis", *J. Clin. Microbiol.* **27**:2723). Field inversion gel electrophoresis was carried out using a Dnastar Pulse (Dnastar, USA) apparatus. Forward and reverse pulses were set at 0.33 s and 0.11 s at the beginning of the run and 60 s and 20 s (or 30 s and 10 s) at the end of the run depending on the molecular weight zone to be expanded. The run time was set at 36 h, the voltage used was 100 V and producing about 325 mA and temperature was maintained at 18°C. Lambda concatamers were used as molecular weight markers. At the end of the run, the gels were stained with ethidium bromide, photographed under UV light and transferred onto Nylon membranes according to Maniatis T. et al., 1982, "Molecular cloning: a laboratory manual" Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. 545 pp.

## RESULTS

### 1. Cloning of the 85-C gene of *M. tuberculosis* :

- Since no specific probe or monoclonal antibody was available to detect specifically an 85-C or related antigen which was expected to bear extensive homology to gene 85-A and gene 85-B this screening required the development of a new procedure. The strategy used was based on the PCR amplification of a 245 bp DNA fragment coding for amino acids 18-98 of the mature antigen 85-A chosen because it is surrounded at both ends by highly conserved DNA sequences when the sequences of antigen A and B are aligned (see primers P78 and P79 in Fig. 2A). It was thus speculated that an equivalent homology might exist with the sequence of antigen 85-C in the same region.

- From *Mycobacterium bovis* BCG genomic DNA a 245 bp DNA fragment was readily obtained. The latter was purified and subcloned in a Bluescribe M13+ vector after digestion with *Eco*RI. About 80 recombinant plasmid containing colonies were tested by plating on nylon filters and hybridized in stringent conditions with a labeled synthetic oligonucleotide recognizing either sequence 85-A (5'-TCGCCCCGCCCTGTACCTG) or sequence B (5'-TCACCTGCGGTTTATCTG) within the PCR amplified fragment (see Fig. 2A). Several clones that hybridized with each probe were sequenced and the sequences were all identical to sequence 85-A in the clones hybridizing with probe A. In those hybridizing with probes 85-B, two kinds of sequences were found : either the 85-B sequence (Matsuo K. et al., 1988, "Cloning and expression of the *Mycobacterium bovis* BCG gene for extracellular  $\alpha$ -antigen" *J. Bacteriol.* **170**:3847) or a new 85-B-related sequence described in Fig. 2A. The latter presents a marked sequence divergence covering 24 nucleotides which is totally distinct from sequence A and B (Fig. 2) (The homology to sequence B is only 40% in this region). Assuming these inserts might represent an amplified fragment of the 85-C gene and that this 24 nucleotide sequence is characteristic of the putative 85-C gene an oligonucleotide probe (probe 85-C) based on this sequence was synthesized.

The latter probe was labeled with  $^{32}$ P and used to screen a collection of 24  $\lambda$ gt11 recombinant phages that were selected in our *M. tuberculosis* and *Mycobacterium bovis* BCG  $\lambda$ gt11 libraries by hybridization with a 800 bp *Hind*III DNA fragment of the previously cloned gene 85-A (Borremans L. et al., 1989,

"Cloning, sequence determination and expression of a 32-Kilodalton protein gene of *Mycobacterium tuberculosis*" Infect. Immun. 57:3123).

Among those phages, those containing the "B" oligonucleotide sequence were further selected by analytical PCR assay using probe B (sense) and oligonucleotide P78 as antisense. This small collection of "85-B-related"  $\lambda$ gt11 recombinant was then hybridized with the described oligonucleotide 85-C and one hybridizing  $\lambda$ gt11-M. *tuberculosis* recombinant was retained, characterized by restriction mapping and sequenced.

## 2. Sequence of the 85-C gene :

The 1211 nucleotide sequence derived from various sequenced fragments is represented in Fig. 1. The DNA sequence contains a 1,020 bp long open reading frame, starting at position 150 and ending with a TGA codon at position 1170. The common NH2 terminal amino acid sequence of the antigen 85 proteins, Phe-Ser, Arg-Pro-Gly-Leu (De Bruyn J. et al., 1987, "Purification, partial characterization and identification of a 32 kDa protein antigen of *Mycobacterium bovis* BCG" Microb. Pathogen. 2:351) could be located within this open reading frame from the nucleotide sequence beginning with a TTC codon at position 288 (Fig. 1). Therefore the DNA region upstream of this sequence is expected to code for a signal peptide required for the secretion of this antigen. The mature protein consists of 284 amino acid residues corresponding to a calculated molecular weight of 32,021.

Interestingly, the N-terminal sequence of the mature protein contains the entire 28 amino acid sequence (phe-ser-arg-pro-gly-leu-pro-val-glu-tyr-leu-gln-val-pro-ser-ala-ser-met-gly-arg-asn-ile-lys-val-gln-phe) described by Wiker H.G. et al., 1990, "Evidence for three separate genes encoding the proteins of the mycobacterial antigen 85 complex" Infect. Immun. 58:272, and which differs only from the common 85-B and 85-A sequence by an alanine instead of a proline in position 16 of the mature protein. Two ATG codons were found to precede the TTC phenylalanine codon at nucleotide position 288 (Fig. 1) in the same reading frame. Use of these two ATG would lead to the synthesis of signal peptides of either 21 or 46 amino acid residues (the latter situation has been represented in Fig. 1 for reasons indicated below).

The base composition of antigen 85-C gene was identical to that of the 85-A gene with an overall G-C composition of 64.57% and a strong preference for G or C in codon position 3 (average 85%). In contrast to antigen 85-A and 85-B that contain 3 cysteines, the sequence of antigen 85-C shows a single cysteine residue at position 254. In fact the two substituted cysteines are located in the region of the mature 85-C protein which contains the largest divergent sequence bloc (Fig. 2B) (SQSNGQNY) (The corresponding DNA sequence was used to synthesize the oligonucleotide probe "C" (see above)). Not surprisingly, this hydrophilic region is also the most divergent when the hydropathy plots of the 3 antigens are compared and thus could be either a variable "epitope" of all 85-antigens and/or a characteristic epitope of antigen 85-C since it was also found in antigen 85-C from *M. bovis* BCG.

Another characteristic feature of antigen 85-C is the presence of the unusual hydrophobic repetitive proline alanine motive PPAAPAAPAA at the carboxy-terminal of the molecule.

## 3. Hydropathy pattern :

The hydropathy pattern of *M. tuberculosis* 85-C antigen was determined by the method of Kyte and Doolittle (Kyte J. et al., 1982, "Simple method for displaying the hydropathy character of a protein" J. Mol. Biol. 157:105). The octapeptide profiles were compared to antigen 85-A and 85-B (Fig. 3). As anticipated from the amino acid sequences, the patterns are roughly similar for the three antigens except for some major differences at region 84-92 and in the carboxy-terminal part of the three proteins.

## 4. Sequence homologies :

DNA sequences from antigen 85-A (Borremans L. et al., 1989, "Cloning, sequence determination and expression of a 32-Kilodalton protein gene of *Mycobacterium tuberculosis*" Infect. Immun. 57:3123 ; De Wit L. et al., 1990, "Nucleotide sequence of the 32 kDa-protein gene (antigen 85A) of *Mycobacterium bovis* BCG" Nucl. Ac. Res. 18:3985), 85-B (Matsuo K. et al., 1988, "Cloning and expression of the *Mycobacterium bovis* BCG gene for extracellular  $\alpha$ -antigen" J. Bacteriol. 170:3847 ; Maniatis T. et al., 1982, "Molecular cloning: a laboratory manual" Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. 645 pp) and 85-C were aligned. An alignment of the three DNA sequences is shown in Fig. 2A. At the DNA level, the homology is maximal between the regions coding for the 3 mature proteins. In this region, the homology between A and B is 77.5% whereas it reaches only 70.8% between the coding regions of genes A and C

and 71.8% between B and C, respectively. Beyond nucleotide 1389 of sequence 85-A and upstream nucleotide position 475 (i.e. within the signal sequence and promoter region) there is practically no homology between the 3 sequences. No significant homology was detected to other DNA sequences present in latest release of GenBank-EMBL.

Homologies at the amino acid level, are presented in the alignment (Fig. 2B) and summarized in Table 1, indicating again a higher homology between sequences A and B (80.4%) than between B/C or A/C.

Other comparisons between the 85-C antigen and the entire SwissProt-NBRF data bank failed to detect any significant homologies to the 85-C antigen amino acid sequence. As for the 85-A antigen, the 85-C sequence does not contain the RGD motif of fibronectin binding proteins nor does it share any homology to the known fibronectin receptors or to the fibronectin binding protein from *Staphylococcus aureus*.

Comparison of the partial DNA sequences of the 85-C gene of the *M. bovis* BCG strain 1173P2 with that of the *M. tuberculosis* strain shows 83.8% homology at the DNA level (Fig. 2A). The region corresponding to the oligo C is entirely conserved. Major differences in the region of the 85-C gene DNA sequence corresponding to nucleotides 815 to 829 of antigen 85-A result in 5 amino acid changes (Fig. 2B). Interestingly the DNA sequence of BCG 85-C within this divergent region is 100% homologous to the sequence 85-B (Fig. 2A).

#### 5. Genome characterization :

In order to confirm the existence of different genes coding for the antigen 85 complex *M. bovis* BCG genomic DNA was digested with SphI, EcoRI and KpnI and the distribution of radioactive signals was examined in Southern blot after hybridization with three specific oligonucleotide (A, B, C) probes (see Material and Methods and Fig. 2A). Three clearly distinct patterns were obtained confirming the specificity of these probes. Similar type specific profiles could be obtained with three random priming labeled DNA restriction fragments (probe 85-A, 230 bp, 85-B, 400 bp, 85-C 280 bp) which were selected within the promoter signal sequence of the three DNAs (Fig. 2A and 4A). With these three DNA restriction fragments, additional weak bands are also observed which clearly correspond to cross hybridization of the probes to the other two genes. With probe 85-C an additional KpnI fragment was observed that does not hybridize to the C-oligonucleotide probe. This probably indicates that the corresponding KpnI site is located upstream of this gene. Furthermore the size of the observed restriction fragments are not always exactly as expected from the restriction maps of the corresponding cloned genes. These discrepancies probably correspond to some minor sequence differences (restriction polymorphism) possibly in non coding DNA regions (outside of the DNA coding for the antigen 85) between strain of *M. bovis* BCG and the *M. bovis* BCG (strain Tokyo) and *M. tuberculosis* respectively.

#### 8. Pulse field analysis of *M. tuberculosis* genomic DNA :

When the largest available 85-A clone BY-5 was hybridized (Fig. 4A) with oligonucleotide B, no positive signal was detected whereas oligonucleotide A gave a positive hybridization (not shown). This indicates that gene B is not located within 2-2.5 kb of the 5' and 4.0 kb of the 3' border of gene A (Fig. 4A). To confirm and extent this result, pulse-field separated DraI-digested *M. tuberculosis* genomic DNA was further hybridized with three specific DNA probes 85-A, 85-B and 85-C in stringent conditions.

Eight strains of *M. tuberculosis* were compared showing six different patterns, three of which are illustrated in Fig. 5. For most strains examined the three probes hybridized to fragments of different sizes. For instance, in *M. tuberculosis* H37Ra, the size of the DraI fragments hybridizing with probes 85-A, B and C were about 242 kb, 212 kb and 245 kb for strain H37Ra, 403 kb, 212 kb and 104 kb for strain H37Rv and 365 kb, 104 kb and 153 kb for strain "1025". Although various strains show some restriction fragment length polymorphism with restriction endonuclease DraI, the simplest interpretation of these results is that the three antigen 85 genes are distantly located (> 100 kb) within the mycobacterial genome.

#### Claims

##### 1. Nucleic acid

- containing a nucleotide sequence extending from the extremity constituted by the nucleotide at position (1) to the extremity constituted by the nucleotide at position (149) represented on Figure 1,
- or containing one at least of the nucleotide sequences coding for the following peptides or polypeptides :

- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (-1) represented on Figure 1, or
- the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (-1) represented on Figure 1, or
- SQSNGQNY, or
- PMVQIPRLVA, or
- GLTLRTNQTPROTYAADGGRNG, or
- PPAAPAAPAA,
- or containing nucleotidic sequences :
  - hybridizing with the above-mentioned nucleotide sequences, or their complements,
  - complementary to the above-mentioned nucleotide sequences, or
  - which are the above-mentioned nucleotide sequences wherein T can be replaced by U.

2. Nucleic acid according to claim 1, containing the nucleotide sequence coding for the following peptide :

SQSNGQNY

and liable to hybridize with the following nucleotide sequence :

CGGCTGGGAC(or T)ATCAACACCCCGGC

and liable to hybridize neither with

GCCTGCGGCAAGGCCGGTTGCCAG

nor with

GCCTGCGGTAAGGCTGGCTGCCAG

nor with

GCCTGCGGCAAGGCCGGCTGCACG.

3. Nucleic acid according to anyone of claims 1 or 2, containing an open reading frame coding for a polypeptide

- liable to react selectively with human sera from tuberculosis patients and particularly patients developing an evolutive tuberculosis,
- or liable to be recognized by antibodies also recognizing the amino acid sequence extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (284) represented on Figure 1,
- or liable to generate antibodies recognizing the amino acid sequence extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (284) represented on Figure 1.

4. Nucleic acid according to anyone of claims 1 to 3, containing an open reading frame and coding for a mature polypeptide of about 30 to about 35 kD and containing a sequence coding for a signal sequence.

5. Nucleic acid according to claim 1, containing a nucleic acid coding for the following polypeptides :

- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (-1) represented on Figure 1, or
- the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (-1) represented on Figure 1, or
- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (284) represented on Figure 1, or
- the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (284) represented on Figure 1, or
- the one extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (284) represented on Figure 1.

6. Nucleic acid sequence according to anyone of claims 1 or 2, containing a nucleic acid coding for a polypeptide sequence extending from the extremity constituted by amino acid at position (18) to the extremity constituted by amino acid at position (98) represented on Figure 2B, on the fifth line.

7. Polypeptide coded by the nucleic acids of anyone of claims 1 to 6.

8. Polypeptide according to claim 7, containing in its polypeptide chain one at least of the following amino

acid sequences :

- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (-1) represented on Figure 1,
- or the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (-1) represented on Figure 1,
- SQSNGQNY,
- PMVQIPRLVA,
- GLTLRTNQTPRDTYAADGGRNG,
- PPAAPAAPAA

9. Polypeptide according to claim 8, containing in its polypeptide chain the following amino acid sequence:

SQSNGQNY

and the amino acid sequence

GWDINTPA

and containing not the amino acid sequence

ACGKAGCQ

and not the amino acid sequence

ACGKAGCT

10. Polypeptide according to anyone of claims 8 or 9, liable to react selectively with human sera from tuberculosis patients and particularly patients developing an evolutive tuberculosis, or liable to be recognized by antibodies also recognizing the polypeptidic sequence extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (294) represented on Figure 1, or liable to generate antibodies recognizing the polypeptidic sequence extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (294) represented on Figure 1.

11. Polypeptide according to anyone of claims 8 to 10, of about 30 to about 35 kD and preceded by a signal peptide.

12. Polypeptide according to claim 8, containing in its polypeptide chain one at least of the following amino acid sequences :

- the one extending from the extremity constituted by amino acid at position (1) to the extremity constituted by amino acid at position (294) represented on Figure 1, or
- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (294) represented on Figure 1, or
- the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (294) represented on Figure 1, or
- the one extending from the extremity constituted by amino acid at position (-48) to the extremity constituted by amino acid at position (-1) represented on Figure 1, or
- the one extending from the extremity constituted by amino acid at position (-21) to the extremity constituted by amino acid at position (-1) represented on Figure 1.

13. Polypeptide according to anyone of claims 8 or 9, containing in its amino polypeptide chain the amino acid sequence extending from the extremity constituted by amino acid at position (18) to the extremity constituted by amino acid at position (99) represented on Figure 2B, on the fifth line.

14. Amino acid sequences constituted by a polypeptide according to anyone of claims 7 to 13, and a protein or an heterologous sequence with respect to said polypeptide, said protein or heterologous sequence comprising from about 1 to about 1.000 amino acids, the heterologous protein being advantageously  $\beta$ -galactosidase.

15. Recombinant nucleic acid containing at least one of the nucleotide sequences according to anyone of claims 1 to 6, inserted in a heterologous nucleic acid.

16. Recombinant vector, particularly for cloning and/or expression, comprising a vector sequence, notably

of the type plasmid, cosmid or phage, and a recombinant nucleic acid according to anyone of claims 1 to 6, in one of the non essential sites for its replication.

17. Recombinant vector according to claim 16, containing in one of its non essential sites for its replication necessary elements to promote the expression of polypeptides according to anyone of claims 7 to 13 in a cellular host and possibly a promoter recognized by the polymerase of the cellular host, particularly an inducible promoter and possibly a signal sequence and/or an anchoring sequence.
18. Recombinant vector according to claim 17, containing the elements enabling the expression by *E. coli* of a nucleic acid according to anyone of claims 1 to 15 inserted in the vector, and particularly the elements enabling the expression of the gene or part thereof of  $\beta$ -galactosidase.
19. Cellular host which is transformed by a recombinant vector according to anyone of claims 16 to 18, and comprising the regulation elements enabling the expression of the nucleotide sequence coding for the polypeptide according to anyone of claims 7 to 13 in this host.
20. Cellular host according to claim 19, chosen from among bacteria such as *E. coli*, transformed by the vector according to claim 18, or chosen from among eukaryotic organism, transformed by the vector according to claim 18.
21. Expression product of a nucleic acid expressed by a transformed cellular host according to anyone of claims 19 or 20.
22. Antibody characterized by the fact that it is directed against a recombinant polypeptide according to anyone of claims 7 to 13.
23. Nucleotide probes, hybridizing with anyone of the nucleic acids according to anyone of claims 1 to 6 or with their complementary sequences.
24. Process for preparing a recombinant polypeptide according to anyone of claims 7 to 13 comprising the following steps:
  - the culture in an appropriate medium of a cellular host which has previously been transformed by an appropriate vector containing a nucleic acid according to anyone of claims 1 to 6, and
  - the recovery of the polypeptide produced by the above said transformed cellular host from the above said culture medium.
25. Method for the in vitro diagnosis of tuberculosis in a patient liable to be infected by *Mycobacterium tuberculosis* comprising
  - contacting a biological sample taken from a patient with a polypeptide according to anyone of claims 7 to 13, under conditions enabling an in vitro immunological reaction between said polypeptide and the antibodies which are possibly present in the biological sample and
  - the in vitro detection of the antigen/antibody complex which has been possibly formed.
26. Method for the in vitro diagnosis of tuberculosis in a patient liable to be infected by *M. tuberculosis*, comprising the following steps:
  - contacting the biological sample with an appropriate antibody according to claim 22, under conditions enabling an in vitro immunological reaction between said antibody and the antigens of *M. tuberculosis* which are possibly present in the biological sample and
  - the in vitro detection of the antigen/antibody complex which may be formed.
27. Necessary or kit for an in vitro diagnostic method for tuberculosis in a patient liable to be infected by *Mycobacterium tuberculosis* according to claim 25, comprising
  - a polypeptide according to anyone of claims 7 to 13,
  - reagents for making a medium appropriate for the immunological reaction to occur,
  - reagents enabling to detect the antigen/antibody complex which has been produced by the immunological reaction, said reagents possibly having a label, or being liable to be recognized by a labeled reagent, more particularly in the case where the above mentioned polypeptide is not labeled.

28. Necessary or kit for an in vitro diagnostic method for tuberculosis in a patient liable to be infected by Mycobacterium tuberculosis according to claim 28, comprising

- an antibody according to claim 22,
- reagents for making a medium appropriate for the immunological reaction to occur,
- reagents enabling to detect the antigen/antibody complexes which have been produced by the immunological reaction, said reagents possibly having a label or being liable to be recognized by a label reagent, more particularly in the case where the above mentioned antibody is not labeled.

29. Immunogenic composition comprising a polypeptide according to anyone of claims 7 to 13, in association with a pharmaceutically acceptable vehicle.

30. Vaccine composition comprising among other immunogenic principles anyone of the polypeptides according to claims 7 to 13 or the expression product of claim 21, possibly coupled to a natural protein or to a synthetic polypeptide having a sufficient molecular weight so that the conjugate is able to induce in vivo the production of antibodies neutralizing Mycobacterium tuberculosis, or induce in vivo a cellular immune response by activating M. tuberculosis antigen-responsive T cells.

31. Peptides of claim 7, advantageously used to produce antibodies, particularly monoclonal antibodies and which have the following amino acid sequences (referring to Figure 1) :

38	H <sub>2</sub> N-DGLRAQDDYNGWDINTPAFE-COOH	57
78	H <sub>2</sub> N-TDWTQPSQSNQNYTYKWET-COOH	97
174	H <sub>2</sub> N-ANSMWGPSSDPAWKRNDPMV-COOH	193
204	H <sub>2</sub> N-RIVVYCGNGTPSDLGGDNIP-COOH	223
235	H <sub>2</sub> N-NQTFRDTYAADGCRNGVFNF-COOH	254
250	H <sub>2</sub> N-GVFNFPFNGTHSWPYWNEQL-COOH	269
275	H <sub>2</sub> N-DIQHVLNGATPPAAPAAPAA-COOH	294

32. Mycobacterium bovis BCG vaccine strain transformed by a recombinant DNA sequence comprising a polypeptide according to anyone of claims 7 to 13, and an epitope of a polypeptide sequence heterologous with respect to said polypeptide.

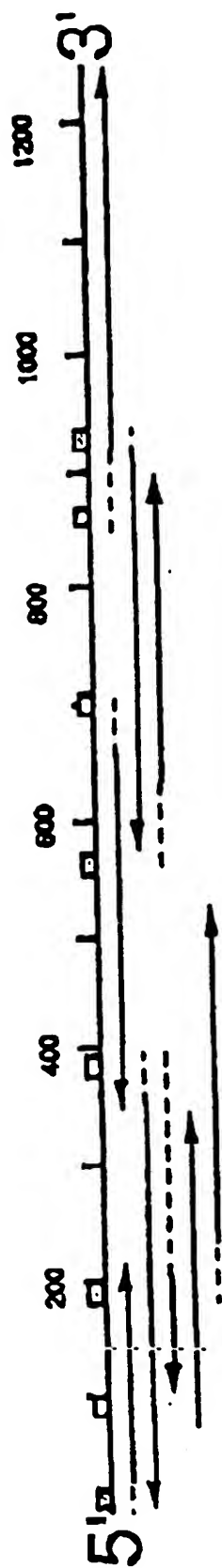


Figure 1



-46	THR	PHE	PHE	GLU	GLN	VAL	ARG	ARG	LEU	ARG	SER	ALA	ALA	THR	THR	LEU	PRO
	ACG	TTC	TTC	CAG	CAG	CTG	CGA	AGG	TTC	CGG	AGC	GCA	GCG	ACA	ACC	CTG	CCG
	ATG	158	158	167	167	176	176	176	176	176	185	185	185	194	194	203	203
-28	ARG	VAL	ALA	ILE	ALA	ALA	MET	GLY	ALA	VAL	LEU	VAL	TYR	GLY	LEU	VAL	GLY
	CGC	GTC	GCT	ATC	CCG	GCT	ATG	GGG	GCT	GTC	CTG	GTT	TAC	GGT	CTG	GTC	GGT
	212	212	221	221	221	230	230	230	230	239	239	248	248	257	257	257	257
-10	THR	PHE	GLY	PRO	ALA	THR	ALA	GLY	ALA	ALA	PHO	SER	ARG	PRO	GLY	LEU	VAL
	ACC	TTC	GGC	GGG	CCG	ACC	CCG	GGC	GCA	TTC	TCT	AGG	CCC	GGT	CTT	CCA	GTC
	266	266	275	275	275	284	284	284	293	293	302	302	311	311	311	311	311
9	GLU	TYR	LEU	GLN	VAL	PRO	SER	ALA	SER	MET	GLY	ARG	ASP	ILE	VAL	GLN	PHO
	GAA	TAT	CTG	CAG	CTG	CCA	TCC	CCG	TCC	ATC	GGC	CCC	GAC	ATC	AGG	GTC	CTC
	320	320	320	320	329	329	338	338	338	347	347	356	356	365	365	365	365
27	GLN	GLY	GLY	PRO	ALA	VAL	TYR	TAC	TAC	CTG	CTC	ASP	GLY	LEU	ARG	ALA	GLN
	CAG	GGC	GGA	CCG	CAC	CCG	GTC	GTC	GTC	CTG	CTC	GAC	GCT	CTG	CGG	GCC	CRG
	374	374	383	383	383	383	392	392	392	401	401	410	410	419	419	419	419

Figure 1 (cont'd)

45 Asp Tyr Asn Gly Trp Asp Ile Asn Thr Pro Ala Phe Glu Glu Tyr Gln Ser  
 GAC TAC AAC GGC TGG GAC GAC ATC AAC ACC ACC GCG GCC TTC GAG GAG GAG TAC TAC 473  
 428  
 63 Leu Ser Val Ile Met Pro Val Gly Gly Gln Ser Ser Phe Tyr Thr Asp Trp  
 GGG TTG TCG TCG GTG ATC ATG ATC CCC GTC GTC GGC GGC CAA TCC AGT TTC TAC ACC GAC TCG 527  
 482  
 72 Tyr Gln Pro Ser Gln Ser Asn Gly Gln Asn Tyr Thr Tyr Lys Trp Gln Thr Phe  
 TAT CAG CCC CCC TCG CAG CAG AGC AAC AAC GGC GGC AAC TAC TAC ACC ACC TAC TAC TCG GAG ACC ACC 561  
 536  
 90 Leu Thr Arg Gln Met Pro Ala Trp Leu Gln Asn Lys Gly Val Ser Pro Thr  
 CTT ACC AGA GAG ATG ATG CCC GGC GGC CTA CAG CAG GCC AAC AAC Lys GGC GGC GTG TCC CCG ACA 635  
 590  
 108 Gly Asn Ala Ala Val Gly Leu Ser Met Ser Gly Gly Ser Ala Leu Ile Leu Ala  
 GGC AAC GCG GCG GTC GGT CTT TCG ATG TCG TCG GGC GGT TCC GCG GCG CTG ATC CTG GCC 689  
 644  
 126 Ala Tyr Tyr Pro Gln Gln Phe Pro Tyr Ala Ala Ser Leu Ser Ser Gly Phe Leu Asn  
 GCG TAC TAC CCG CAG CAG CAG TTC CCG TAC TAC GCC GCC GCG TCG TCG TTG TCG TCG GGC TTC CTC AAC 743  
 698  
 707  
 716  
 725  
 734

Figure 1 (con't 2)

144	Pro	Ser	Gln	Gly	Gly	Tyr	Tyr	Pro	Thr	Leu	Ile	Gly	Leu	Ala	Met	Asn	Asp	Ser	Gly
	CCG	TCC	GAG	GAG	GGC	TGG	TGG	CCG	ACG	CTG	ATC	GGC	CTG	GCG	ATG	AAC	GAC	TCG	GCC
			752			761			770			779				788			797
162	Gly	Tyr	Asn	Ala	Asn	Ser	Met	Tyr	Gly	Pro	Ser	Ser	Ser	Asp	Pro	Ala	Tyr	Lys	Arg
	GGT	TAC	AAC	GCC	AAC	AGC	ATG	TGG	GGT	CCG	TCC	AGC	AGC	GAC	CCG	GCC	TGG	AAG	CGC
			806			815			824			833				842			851
180	Asn	Asp	Pro	Met	Val	Gln	Ile	Pro	Arg	Leu	Val	Ala	Ala	Asn	Asn	Thr	Arg	Ile	Tyr
	AAC	GAC	CCA	ATG	GTT	CAG	ATT	CCC	CGC	CTG	GTC	GCC	GCC	AAC	AAC	ACC	CGG	ATC	TGG
			860			869			878			887				896			905
198	Val	Tyr	Cys	Gly	Asn	Gly	Thr	Pro	Ser	Asp	Leu	Gly	Gly	Gly	Asp	Asn	Ile	Pro	Ala
	GTC	TAC	TGC	GGT	AAC	GGC	ACA	CCC	AGC	GAC	CTC	GGC	GGC	GAC	GAC	AAC	ATA	CCG	GCC
			914			923			932			941				950			959
216	Lys	Phe	Leu	Gln	Gly	Leu	Thr	Leu	Arg	Thr	Asn	Gln	Thr	Phe	Arg	Arg	Asp	Thr	Tyr
	ANG	TTC	CTG	GAA	GGC	CTC	ACC	CTG	CGC	ACC	AAC	CAG	CAG	ACC	TTC	CGG	GAC	ACC	TAC
			968			977			986			995				1004			1013
234	Ala	Ala	Asp	Gly	Gly	Arg	Asn	Gly	Val	Phe	Asn	Phe	Pro	Pro	Pro	Asn	Gly	Thr	His
	GCG	GCC	GAC	GGT	GGA	CGC	AAC	GGG	GTC	TTT	AAC	TTC	CCG	CCC	ACC	AAC	GGA	ACA	CAC
			1022			1031			1040			1049				1058			1067

Figure 1 (con't 3)

252 Ser TTP Pro Tyr TTP Asn Glu Gln Leu Val Ala Met Lys Ala Asp Ile Gln His  
 TCG TGG CCC TAC TGG AAC GAG GAG CAG CAG CTG GTC GCC ATG ATG AAG GCC GAT GAT GAT GAT CAT  
 1076 1085 1094 1103 1112 1121  
 270 Val Leu Asn Gly Ala Thr Pro Pro Pro Ala Ala Pro Ala Ala Ter  
 GTG CTC AAC GGC GCG ACA CCC CCC GCG GCC GCG CCT GCT GCG CCG GCC GCG TCGGCCA  
 1130 1139 1148 1157 1166 1176  
 GCAGCCAGCATCGGCAGCAGCCGACGCGCCAGCGCG  
 1185 1194 1202 1211

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Figure 1 (con't 4)

	10	20	30	40
85A-TUB	★	★	★	★
85B-BCG				
85B-KAN				
85C-TUB				
85C-BCG				

TGACCGCACCGGATACGTTGCGGCAGGCATCTGGCTGGCGG-TGGTT  
 -----  
 .TT.--A.T..T.T..TAC-----TC.CC.C..C..CC.TC.G  
 -----  
 -----TG.C....C.A.C...  
 -----  
 -----

	50	60	70	80	90
85A-TUB	★				★
85B-BCG					
85B-KAN					
85C-TUB					
85C-BCG					

	100	110	120	130	140
85A-TUB	★	★	★	★	★
85B-BCG					
85B-KAN					
85C-TUB					
85C-BCG					

**Figure 2A**

**Figure 2A (con't 1)**

85A-TUB  
85B-BCG  
85B-KAN  
85C-TUB  
85C-BCG

300      310      320      330      \* PstI

GTGTGTTGACT-ACACG---AGCACTGCCGGGCCAGCGCTGCAGTC  
..C...-.C...-.C...-.C...-.C...-.C...-.C...-.C...-A  
CA.T..CCCT..C...GTA..TT...TAGT...TAGT...TAGT...A  
....G.A...-.C...-.C...-.C...-.C...-.C...-.C...-A  
-----

85A-TUB	340 *	350 *	360 *	370 *	380 *
85B-BCG	TGACCTAATTCAGGATGCCCCCAACATGCATGGATGCGTTGAGATGAG-				
85B-KAN	C..GAC..A.....GGG...CA.G.-----CA				
85C-TUB	.....C.C...AG..C.T.C.G.-----AAG.G.CA.G.-				
85C-BCG	...G.TC.....A-----A-				

	390	400	410	420	430
	↑	★	★	★	★
85A-TUB	GATGAGGAAGCAAGATATGCAGCTTGTGTGACAGGGTTCGTGGCGCGGTCA				
85B-BCG	..C.T.A.CC.A....T.CG.....GG..				
85B-KAN	T....CA..C.TG..CGG.A.A.....C.GG.....G.....				
85C-TUB	C.G.T.C.....T.G..GA...A.CG.				
85C-BCG	-----				

**Figure 2A (con't 2)**

	440	450	460	470	480
85A-TUB	CGGGTATGTCGGCTCGACTCGTGT	CGGGGCGCTCGGCGGCCCTAGTG			
85B-BCG	-----C...T.GA..A...	CA.G.CA.CG..T.TAG.CC.T			
85B-XAN	-----A..C.TT..G..C.G..C...	CT.CT.....T			
85C-TUB	.AACC...C...C.CG.G.CTA..C...	TA.G..G..T.T..G..T			
85C-BCG	-----	-----			

	490	500	510	520	530	
	*	*KpnI	*	*	*	↓
85A-TUB	TCGGGTCTGGT	CGCGCGT	CGGTGGC	ACGGCG	ACCGGGGG	CAATTTTC
85B-BCG	C.....G	GCTT.C	.C.AG.	.A.....C	.G.....C	.C.
85B-KAN	C.T..C.....	.ACT.C	.C.AG.	.A.....A	.G.....C	.C.
85C-TUB	.AC.....	[.TA.]T	.C.GC.	.C.....C	.C.....C	.C.
85C-BCG						

[illegible]

**Figure 2A (con't 3)**



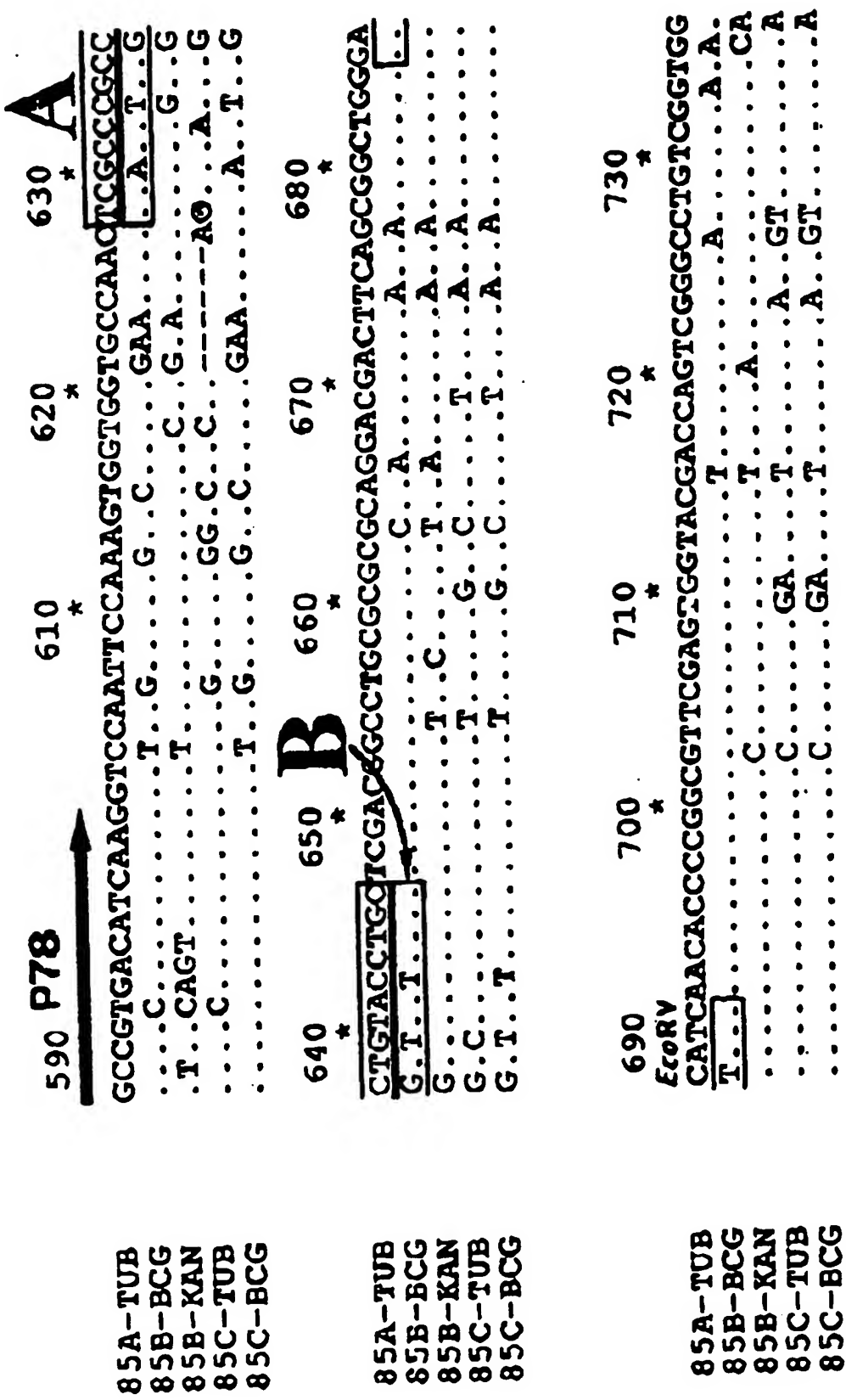


Figure 2A (con't 4)

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85A-TUB	740 *	750 *	760	770 *	780 *
85B-BCG	TCATGCCGGTGGTGGCCAGT	CTAAGCTT	CTACTCCGACTGGTACGCCC		
85B-KAN	.....C..C..G.....	.....C.....	.....AG.....	.....AGC..G	
85C-TUB	.....C..C..A.....	.....C..T.....	.....AGT.....	.....AGC..G	
85C-BCG	.....C.....	.....A..C..T.....	.....A.....	.....T.....	
	.....C.....	.....A..C..T.....	.....A.....	.....T.....	
	790 *	800 *	810 *	P79	830 *
85A-TUB	GCCTGGGCAAGGCCGGT	TGCCAGACTTACAAGTGGGAGACCTT	CTCTGAC		
85B-BCG	.....T.....	.....T..C.....	.....A...C.....		
85B-KAN	.....C.....	.....C...AC.....	.....C.....		
85C-TUB	T.GCAGA.....	C.G.CAGAA.T.C	.....T.....		
85C-BCG	T.GCAGA.....	C.G.CAGAA.T.C	.....T.....		
	840 *	860 *	870 *	880 *	
85A-TUB	CAGCGAGCTGCCGGGTGGCTGCAGGCCAACAGGCACGTCAAGCCACCG				
85B-BCG	.....CAA...T..TCC.....	.....GC...G.....			
85B-KAN	.....CAA.....	TCC..G..C..AGT.....			
85C-TUB	...A...A...C.CC.....	.....A.GG...GTCC..G..A.			
85C-BCG	-----	-----	-----	-----	

Figure 2A (con't 5)

85A-TUB	890 *	900 *	910 *	920 *	930 *
85B-BCG	GAAGCGCGTCGTCGGTCTTTCGATGGCTGCTTCTTCGGCGCTGACGCTG				
85B-KAN	.C....T.CAA...CT.G.....C.GC..G....AA...TCT..				
85C-TUB	.....CG.....CA.C.....C.GC.TG....C....TC...				
85C-BCG	.C.A...G.CG..G.....T.G.GCGG...C.....TC...				
85A-TUB	940 *	950 *	960 *	970 *	980 *
85B-BCG	GCGATCTATCACCCCGCAGCAGTTCGTCTACGGGGAGCGATGTCGGGCGCT				
85B-KAN	..CGC...C.....A.....C..CT..C.....C...				
85C-TUB	T.CG...C.....G.....A.....TT..T.....C...				
85C-BCG	..CGCG..CT...G.....CCG.....C.CGT..T.....T.				
85A-TUB	990 *	1000 *	1010 *	1020 *	1030 *
85B-BCG	GTTGGACCCCTCCAGCGGATGGGTCCACCCCTGATCGGCCCTGGCGATGG				
85B-KAN	.C.....T....G.....G.....C.....C.....				
85C-TUB	.A.....G.....G.....GT..T.....T.....				
85C-BCG	CC.CA...G...G...GCTG.T.G..G..G.....A				

Figure 2A (con't 6)

85A-TUB	1040	1050	1060	1070	1080
85B-BCG	*	*	*	*	*
85B-KAN	GTGACGCTGGCGGCTACAAGGCCTCCGACATGTGGGGGCCGAAGGAGGAC				
85C-TUB	.....C.....T.....G.A.....T..CTC.AGT...				
85C-BCG	.....C..T..T.....G.....A..CTC.AGT...				
	AC...T.G....T....C...AA.AG.....T...TCCAGC...				
	-----				
85A-TUB	1090	1100	1110	1120	1130
85B-BCG	*	*	*	*	*
85B-KAN	CCGGCGTGGCAGCGCAACGACCCGCTGTTGAACGTGCGGAAGCTGATCGC				
85C-TUB	.....A..G.....TAC.CA.C.GA..CCC.....G....				
85C-BCG	.....A..C.....T.....TC.C..C..A.TCC.G.....G....				
	.....C...A.....AA..G.TC.GA.TCCCCGC...G....				
	-----				
85A-TUB	1140	1150	1160	1170	1180
85B-BCG	*	*	*	*	*
85B-KAN	CAACAACACCCGCGTCTGGGTGTACTGCGGCAACGGCAAGCCGTCGGATC				
85C-TUB	A.....GC.A.....T..T.....G.....CC...AAC..GT				
85C-BCG	.....C.G...A.C.....T.....CC.....C..GT				
	.....GA.....T.....CA..CAGC..C..				
	-----				

Figure 2A (con't 7)

85A-TUB	1190	1200	1210	1220	1230
85B-BCG	★	★	★	★	★
85B-KAN	TGGGTGGCAACACCTGCCGGCCAAAGTTCTCGAGGGCTTCGTGCGGACC				
85C-TUB	....C..TGC....A.A..C...G....T.G...AA....T..T.G.				
85C-BCG	....C..TGC....TG.T....G.A....G....AA....T..C.G.				
	.C..C...G....A.A....G....G..A...C..ACC.T.CG.				
	-----				
85A-TUB	1240	1250	1260	1270	1280
85B-BCG	★	★	★	★	★
85B-KAN	AGCAACATCAAGTTCCAAAGACGCCCTACACGCCGGTGGCGG-----CCACA				
85C-TUB	....C.G....G...T..G....GC...CG....G....				
85C-BCG	....C.G..A....G....G....CG....GCGG....				
	.C....CAG.CC...GG...A....GCG...AC..T...A.G..				
	-----				
85A-TUB	1290	1300	1310	1320	1330
85B-BCG	★	★	★	★	★
85B-KAN	ACGGCGTGTTCCGACTTCCCGGACAGCGGTACGCCACAGCTGGGAGTACTGG				
85C-TUB	....C....A....CC...A...C....				
85C-BCG	....C....A..GGAC.C..A...A....				
	....G....TA....CC..A...A..A..TCG...CCC....				
	-----				

Figure 2A (con't 8)

85A-TUB  
85B-BCG  
85B-KAN  
85C-TUB  
85C-BCG

1340 \* 1350 \* 1360 \* 1370 \*  
GGCGGCAGCTCAACGGCTATGAAGCCCGACCTGCAACGGGCA--CTG---  
.....T.....C.....GGT.....GA.TT.G--T.A---  
.....C.....GGT.....C.GC.....  
AA..A.....GGT..C.....TA.C..G.AT.TG--..CAAC  
-----

85A-TUB  
85B-BCG  
85B-KAN  
85C-TUB  
85C-BCG

1380 \* 1390 \* 1400 \* 1410 \*  
GGTGCCA--CGCCCAACACCGGGCCCGCCCGCCAGGGC--GCC  
..C.....CGCTGAT..G.....TTGC...TA.T...TTGAC.G.  
..C..G.....A...CCGG...CC..T..TG.G.C..C.....  
-----

85A-TUB  
85B-BCG  
85B-KAN  
85C-TUB  
85C-BCG

1430 \* 1440 \* 1450 \* 1460 \*  
TAG-CTCCGAACAGACACAACA---TCTAGCNCNCGGTGACCCCTGTGGNN  
GG.-A...C...GG.T...T...TC  
A..A.G...TCA..C..GT.GTGTG.TCG..AC.TTGA..G..G..CCGC  
-----G-----G.C..-----  
-----

Figure 2A (con't 9)

	1470	1480	1490	1500	1510
85A-TUB					
85B-BCG					
85B-KAN					
85C-TUB					
85C-BCG					

	1520	*	1530	*
85A-TUB	AGCTACCTGACNNCATGGGTTTG			-----C
85B-BCG	---CG...TTG..GCAC.AG.CCCCCCG			-----G
85B-KAN	G.TG..A.CGACC....C.-G.			-----G
85C-TUB	..-----			-----G
85C-BCG	-----			-----G

**Figure 2A (cont'd 1D)**

### Figure 2B



85A	200	210	220	230	240	250
85B	LNVCGLIANNTRVWVYCCNGCHP	SDLGNNHLP	AKFLEGFV	RTSNIK	EQDAYN	AGGHHGVF
85B-KA	QQIP..V....L.....T.NE...	A.I..E..N...	S..L.....RPA.....A..			
85C	HIDE.V.D...L.I.....T..E...	ADV..E..N...	S..L.....A.....A..			
	VOIPR.V....I.....T.....D.I.....	LTIRT	QT	R.T.A.D..R....		
85A	260	270	280	290		
85B	DPDQSGTHSWEYNGAQLNNAK	PDLOAL	-GATPNT	CPAPOGA		
85B-KA	N..DN.....G...SS..					
85C	-NL.A.....D...G...AG..					
	N..PN.....P.NE..V...A.I.HV.N....	PAA..APA.				

Figure 2B (con't 1)

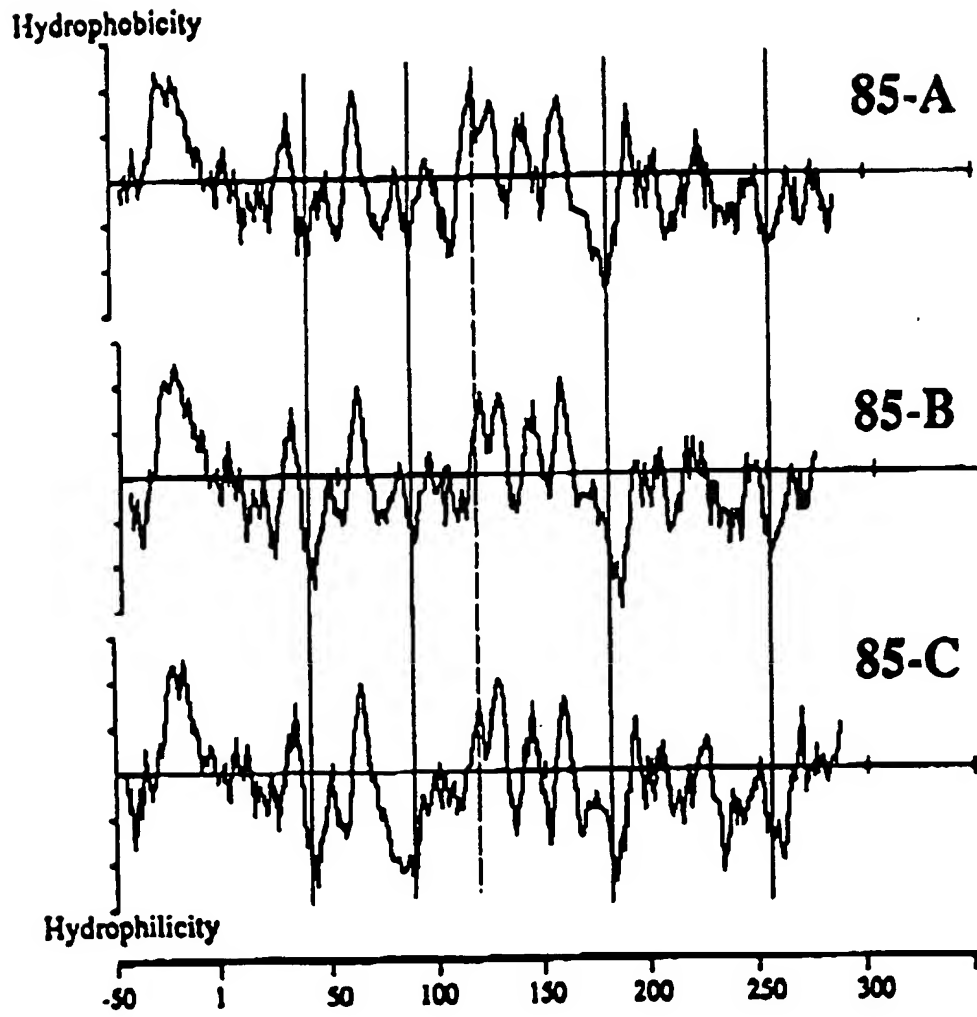


Figure 3

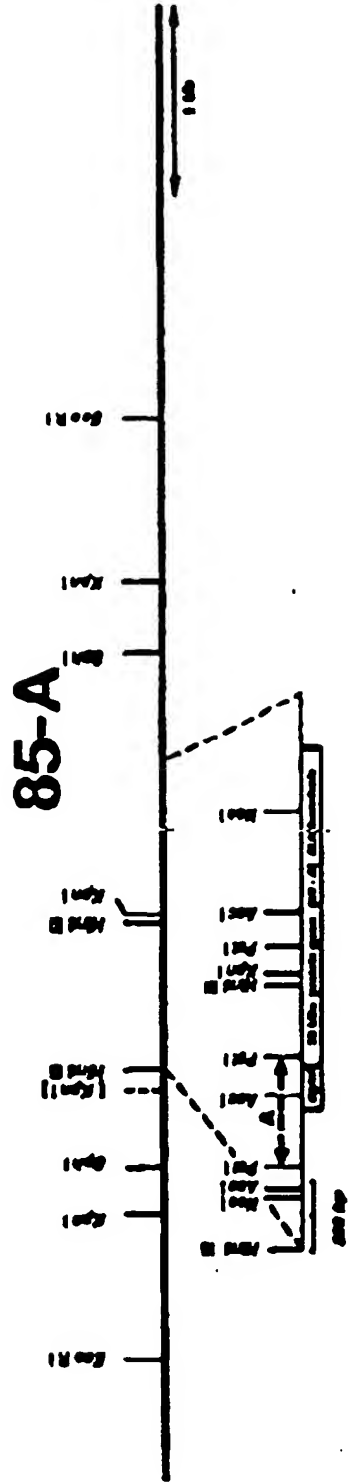
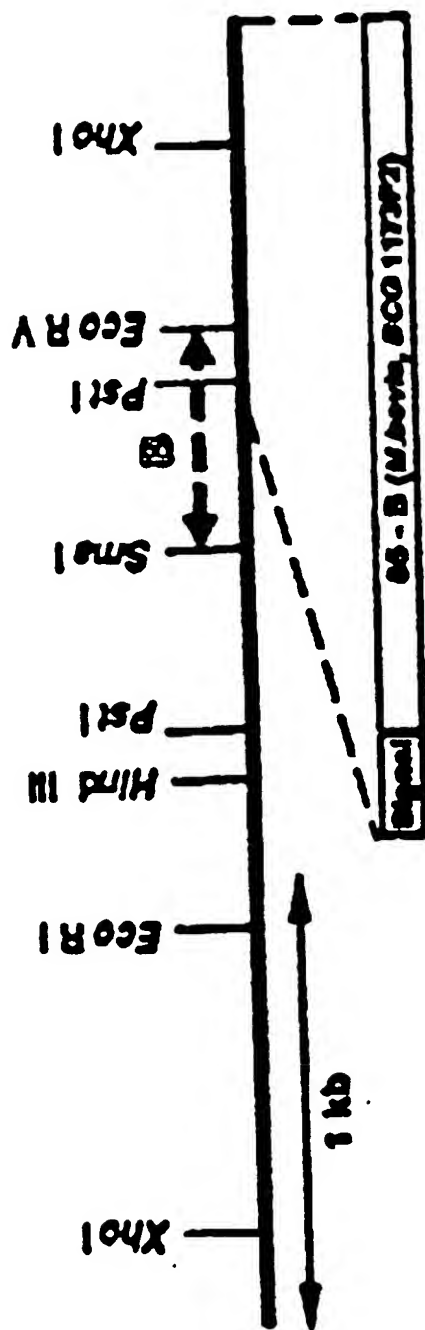


Figure 8A



85-B

Figure 4A (con't 1)

# 85-C

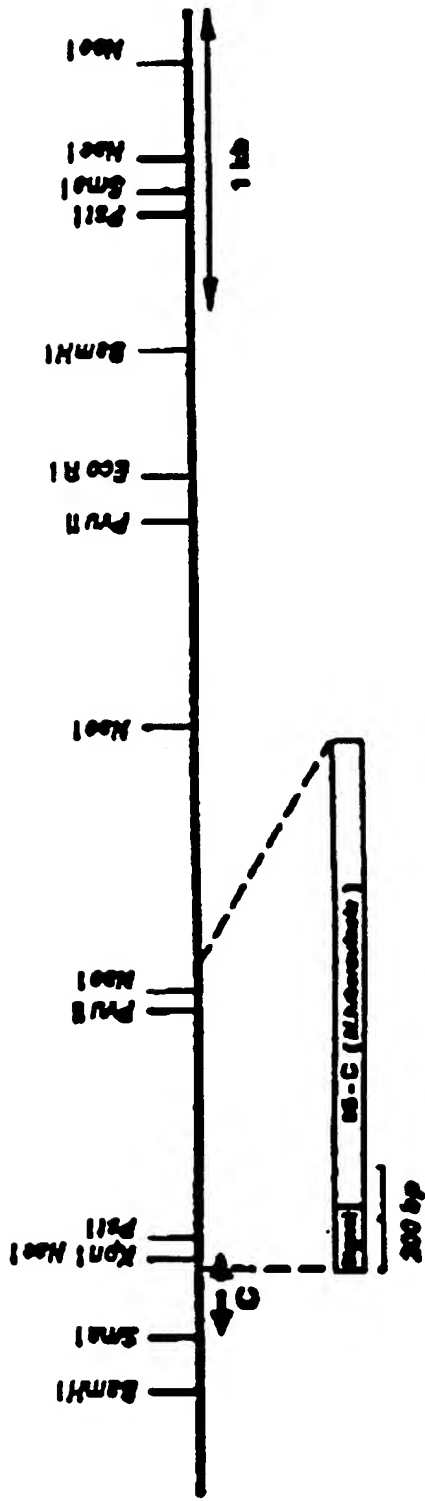


Figure 4A (con't 2)

# 85-A 85-B 85-C

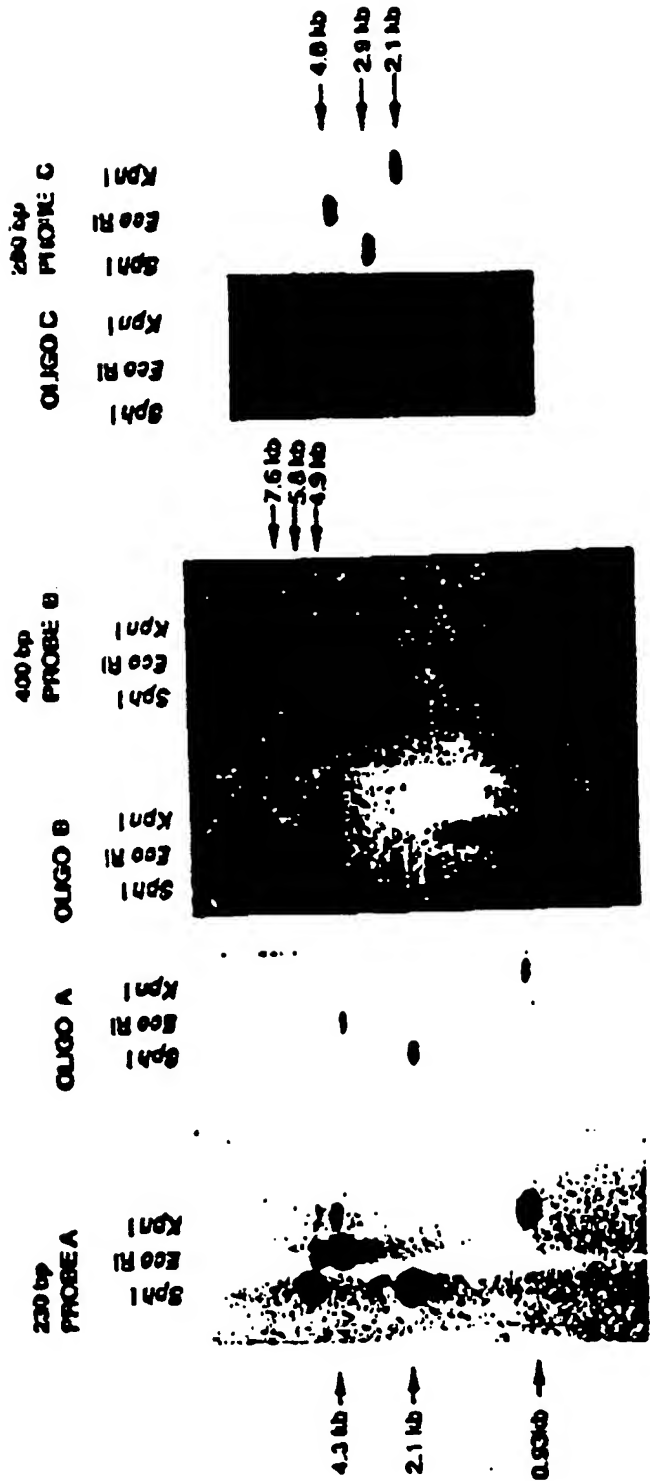


Figure 4B

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Figure 5



European Patent  
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## EUROPEAN SEARCH REPORT

Application Number

EP 91 40 0388

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.)
Y	INFECTION AND IMMUNITY, vol. 59, no. 1, January 1991, WASHINGTON US pages 372 - 382; S. NAGAI ET AL.: 'Isolation and Partial Characterization of Major Protein Antigens in the Culture Fluid of Mycobacterium tuberculosis' " abstract " " table 1 "	1,4, 14-18	C12N15/21 C12Q1/48 G01N33/669 A61K39/04 A61K39/395 C12N15/62 C12P21/08 C12N1/21
Y	WO-A-9 010 701 (COMMONWEALTH SCIENTIFIC AND INDUSTRIAL RESEARCH ORGANISATION) September 20, 1990	1,4, 18-17	
A	" abstract "	19-21,24	
A	WO-A-9 018 876 (INSTITUT NATIONAL DE LA SANTE ET DE LA RECHERCHE MEDICALE (INSERM)) November 1, 1990 " page 18, line 7 - line 25 " " page 21, line 6 - line 16 " " claims 3,47 "	23,25	
			TECHNICAL FIELD SEARCHED (Int. Cl.)
Y	JOURNAL OF BACTERIOLOGY vol. 169, no. 3, March 1987, BALTIMORE, U.S.A pages 1080 - 1088; T. M. SHINNICK: 'The 66-Kilodalton Antigen of Mycobacterium tuberculosis ' " abstract "	14-18	C07K C12N A61K C12P
A	WO-A-8 701 110 (SCRIPPS CLINIC AND RESEARCH FOUNDATION) February 26, 1987 " abstract " " page 6, line 3 - line 15 " " page 8, line 26 - line 29 " " page 13, line 6 - line 22 "	22,26-30	
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 08 OCTOBER 1991	Examiner THIELE U.M.C.H.
CATEGORY OF CITED DOCUMENTS X: particularly relevant if taken alone Y: particularly relevant if combined with another document of the same category A: technological background O: non-written disclosures F: intermediate document		T: theory or principle underlying the invention E: earlier patent document, but published on, or after the filing date D: document cited in the application L: document cited for other reasons S: number of the same patent family, corresponding document	



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